Cleaning and Sanitation Validation: What Does Clean Look Like?

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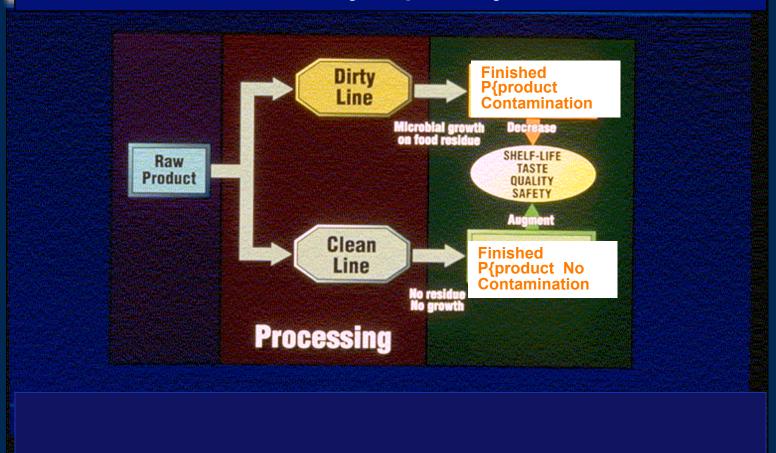
Cleaning and Sanitation Validation: What Does Clean Look Like?

- > Introduction
- Cleaning, Sanitation and Hygiene What does it mean?
- Surface sampling methodology for monitoring and validation of cleaning and sanitation
- Rapid Hygiene Monitoring ATP and non- ATP swabs
- Consideration in Environmental monitoring
- Summary

Food Processors must...

- Prevent contamination
- > Eliminate or reduce the numbers of microorganisms
- Minimize growth and activity of surviving organisms
- Prevent post-processing contamination

Clean line = safety, quality and shelf life



Monitoring Cleaning and Sanitation Efficacy...



- > Why?
- > What?
- > When?
- > How?
- > Who?

WHY CLEANING AND SANITATION ??

- ►It's the Law!
- ➤ It's a good business
- ➤ It's related to product quality and shelf life and consumer satisfaction
- Loss of reputation and Brand protection

WHY CLEANING AND SANITATION ??

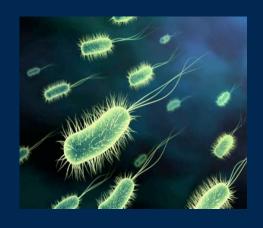






- ➤ 1990s USDA/FDA GMP SSOP Prerequisite to HACCP
- **EEC Council Directives**
 - ➤89/392/on machinery including Agri-food stuff machinery
 - ➤ 93/94 on the hygiene of food stuff 1993
 - ➤ 1990 Food safety Act (U.K)

WHY CLEANING AND SANITATION ??



Its good business

- > Aesthetics and Process Efficacy
- > Equipment Performance/ maintenance
- Spoilage, K.Q and Shelf life
- ➤ Vendor compliance

Universal Emphasis on Hygienic Design and Cleanability of Food Processing Equipment





> Cleaning

Removal of all soil, food product residue, dirt, grease or other objectionable matter.

> Sanitation

All precautions and measures which are necessary in the production, processing, storage and distribution, in order to assure an unobjectionable, sound and palatable product which is fit for human consumption

> Sanitation/ Sanitizing

Application of any effective method or substance to clean surface for the reduction of the bacterial count of pathogens, to a safe and acceptable level and of other organisms to as far as practicable. Such treatments shall nor adversely effect the equipment, the product or the health of the consumer and shall be acceptable to the health authority.

> Disinfection

The reduction, by means of chemical agents and/or physical methods, of the number of micro-organisms in the environment, to a level that does not compromise food safety or suitability.

> Hygiene

Conditions or practices conducive to maintaining health and preventing disease, esp. through cleanliness.

- Cleaning alone, may remove some bacteria. However, it is NOT adequate to reduce the bacterial populations to an acceptable low level
- Clean before sanitation
- Pre-op sanitation

What is a 'Clean' Surface?

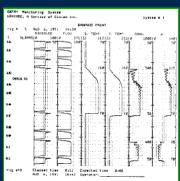
- Physically
 Removal of all soil, and product residue
- Chemically
 Removal of cleaning and sanitizing material by rinsing
- Microbiologically Reduction to an acceptable level of microorganisms

Monitoring Cleaning and Sanitation Efficacy...

Hygiene monitoring – How?

- Visual Inspection
- Checking Cleaning Conditions
- Microbiological Tests
- Rapid Hygiene monitoring Tests (ATP/ non-ATP)







Monitoring: Visual Assessment

- Sensory/visual assessment of equipment/ surfaces
- "Sensitive" or "Worst case" scenario
- Skill and experience of inspector
- > Apparent cleanliness can be misleading!





Surface sampling methodology for monitoring and validation of cleaning and sanitation

Dr. Purnendu C. Vasavada Professor Emeritus- Food Science University of Wisconsin - River Falls

Presented at 2011 IAFP Pre Annual meeting workshop Milwaukee, Wi July 29, 2011



Microbiological Assessment of Food **Contact Surfaces**







- Swab/ Sponge testsRinse tests
- Agar Contact Methods- RODAC
- Direct Surface Agar Plating (DSAP)
- Sticky Tape Technique
- Dye reduction tests
 ATP bioluminescence tests





Sponges and Swabs

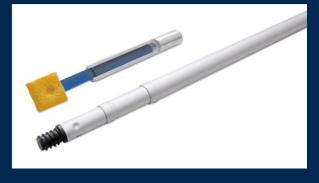












Swab Tests

- > Simple
- ➤ Most common method





- Sample a surface with a sterile cotton swab followed by standard plate count
- > Poor recovery of residual microorganisms
- Poor reproducibility
- ➤ Not suitable for specific "index" organism
- > Improved results with refinements

Total Microbial Counts for a Cannery

Grade	Microbial Counts (cfu/ft²) after swabbing
Satisfactory	0 - 5,000
Fairly satisfactory Unsatisfactory	5,000 - 25,000 > 25,000

¹From Shapton and Shapton (1991)

Rinse Method

- Collection of contamination by rinsing of entire surface followed by standard plate count
- Suitable for small surfaces/area
- > Higher recovery
- More accurate than swab test
- Can be used with membrane filtration









Agar Contact Methods

- > Pressing plates containing agar against a surface followed by incubation and counting
 - **≻**RODAC
- > Petrifilm
- >Hycheck >Monoflex
- Agar Syringe and Agar "Sausage"







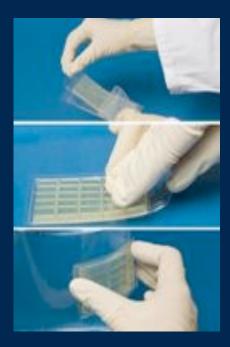


Agar Contact Method





Biotest Contact Slides



Petrifilm application



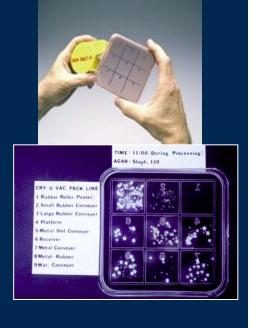




Sticky Tape Technique

- Sterile cellophane tape applied to surface to be tested, reapplied to agar surface followed by incubation and counting
- > CON-TACT-IT®
- Simple, easy, economical
- Similar limitations as agar contact methods
- Potential contamination by the transfer step







Agar Contact Methods – Advantages and Limitations

- Simple
- Commonly used
- Versatile
- Generally suitable for flat surfaces
- Confluent colony/spreaders

Direct Surface Agar Plating (DSAP)

- ➤ In situ assessment of surface contamination by pouring a melted medium, allowing to solidify followed by incubation at room temperature
- Suitable for eating utensils
- Can't use equipment during the test period
- Confluent colony/spreaders

Miscellaneous Methods

- Indicator/Dye reduction
- > in situ use of redox dye(NitroBlue, Tetrazolium)
- Catalase method
- > Epiflurescence Microscopy
- Molecular detection

Rapid Hygiene Monitoring

- > Simple, effective, cleanliness measurement
- > Immediate results
- > "Actionable" results
- >ATP Bioluminescence
- ➤ Non-ATP swabs

Rapid Hygiene Monitoring using ATP Bioluminescence

- > Total ATP Hygiene
- Rapid Real Time
- > Microbial vs. somatic ATP



Some commercially available luminometers

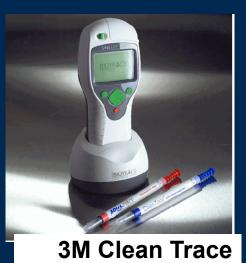
Biocontrol











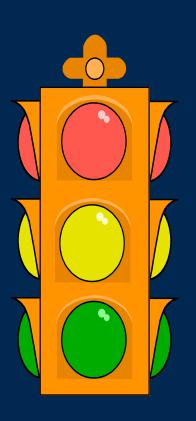
ATP Bioluminescence : Simple, Quantitative Relationship

increase in light (RLU)

increase in ATP levels

increase in organisms or product residues

Interpretation of Results



Red - Yellow - Green Fail - Caution - Pass

Sanitation failure Re-clean prior to start-up

Results trending upward Needs attention

Production may begin Plant is clean

Criteria for Assessing Cleaning Efficacy of Food Contact Surfaces

Cleaning	RLU ¹	Zone units ²
Efficacy		
Efficient	< 5	< 2.0
Moderate/	5 - 50	2.0 - 2.5
poor		
Unclean	> 100	> 3.0

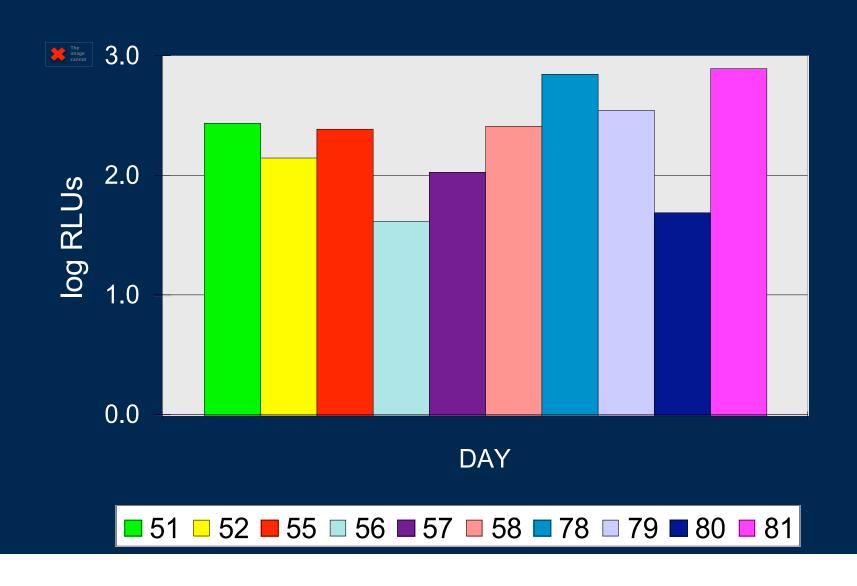
¹Relative Light Units; ²Used with Lightning Luminometer

Hygiene validation – Chees plant surfaces

Surfaces evaluated by	Pass	Fail
Visual Inspection	119	0
Aerobic Plate Counts	47	72
ATP	42	77

Source: Kyrikides et. Al. 1990.

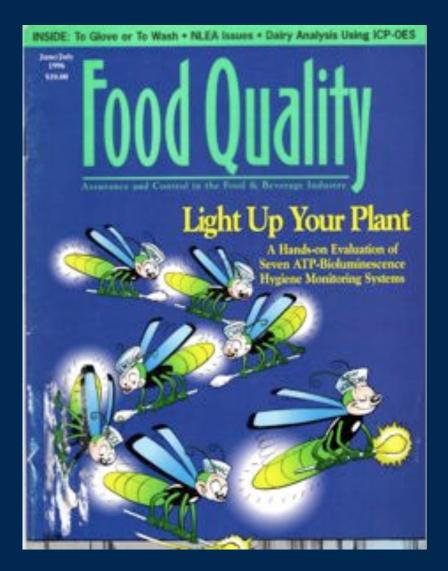
Results from a Particular Day

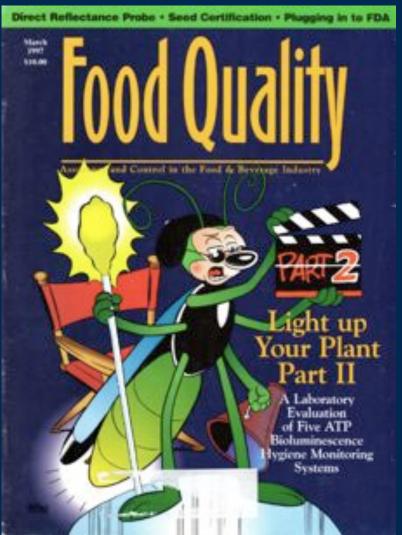


Weekly Sample Data



ATP Luminometer Performance evaluation

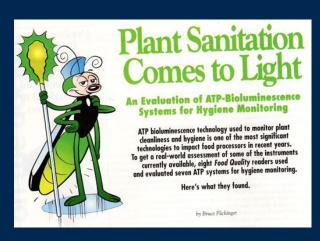




Hygiene Monitoring

	Visual assessment	Microbiological tests	ATP
Rapid	✓	X	✓
Objective	X	✓	✓
Sensitive	X	✓	✓
Detect product residues	✓	X	✓
Simple	✓	✓	✓
		(Lab required)	

Selection Criteria and Considerations



- Sensitivity
- Reproducibility
- Repeatability
- Simplicity
- Instrument
- Reagents and Swabs
- > Training
- Technical Service
- > Industry acceptance

Important considerations

- Surface to be tested
- Method selected
- Lab and skills requirements
- Data handling and interpretation

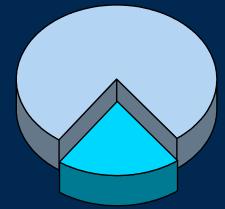
CFU to RLU Correlation

Relative light Units do not equal Colony forming Units



RLU correlates well to CFU with pure culture

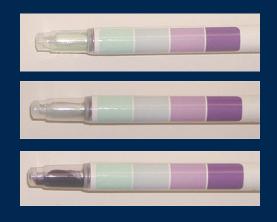
Residual ATP



Microbial ATP Correlation does not occur because in real world environment there is residual ATP as well as microbial ATP.

Non-ATP swabs

- ➤ Detects levels of protein, sugar and other compounds associated with food and microbial contamination.
- Semi-Quantitative darker color/faster color development = more protein present
- > Can detect as little as 50µg of protein



Green - Pass
Grey - Caution
Purple - Fail





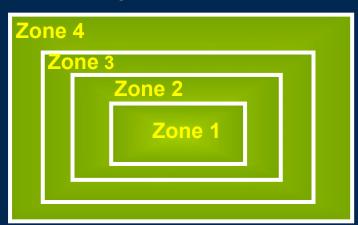
Assessment of cleanliness with AssurSwab/ Swab' N' Check and the ATP methods

Swab 'N'	Check ¹	ATP ²
SS Meat cooking table before washing	4	5300
SS Meat cooking table	1	50-66
after washing Plastic meat processing device	4	4100
after washing w/water alone Plastic meat processing device	2	53, 64
after washing / sanitizing		

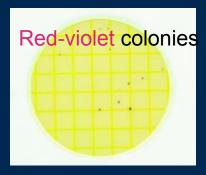
1. Cleanliness level 1-4(1= Clean, 4 = Dirty), 2. RLU

Some Recent Developments

- ➢ Biofilm
- Indicator and Specific Pathogen testing
 - ➤ Listeria spp. or Listeria-like organisms
 - ➤ E. coli, E. coli 0157:H7
- Surveillance of hot spots
- > Zone testing







Biofilms



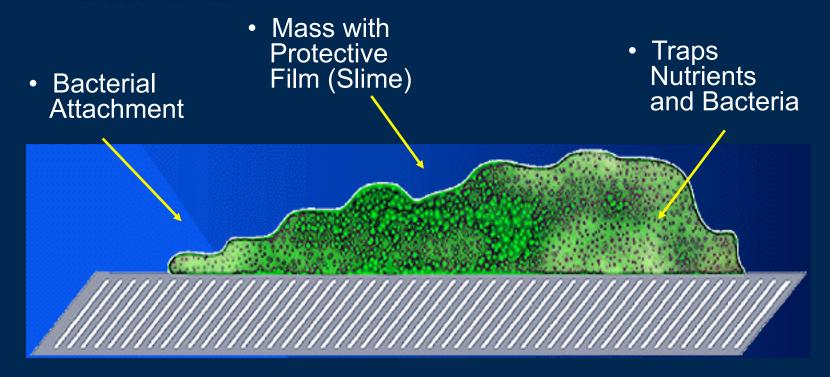






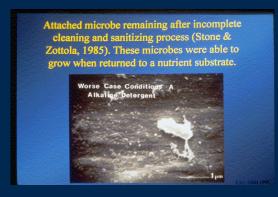


Biofilms



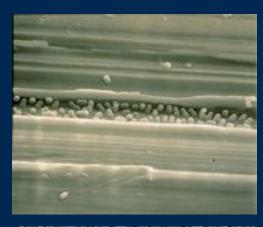
- Prevents Anti-Microbial Action
- Effective Cleaning Required

Microbial Attachments and Biofilms



- Microorganisms e.g. L. m. & Pseudomonas can attach to food contact surfaces and form biofilms.
- Attached microorganisms (*P. fragi*) are not removed or inactivated under less than optimal cleaning and sanitizing of milk pipeline.
- Numbers of attached microorganisms in the biofilm increase to a point where they may resist inactivation by cleaning and sanitizing if intervals between cleaning and sanitation > 8 hr.

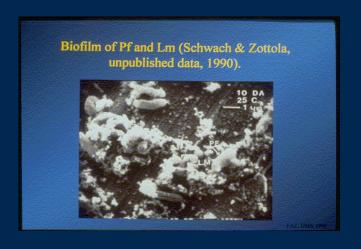
Microbial Attachments and Biofilms

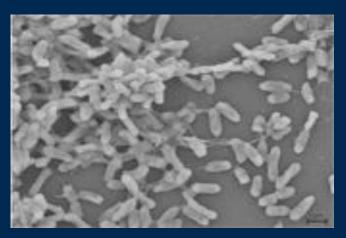




- Attached microorganisms may resist treatment with sanitizer e.g. Microcolonies of L. monocytogenes require 12-20 min. treatment with benzalkonium chloride, vs. 30 sec. for unattached cells.
- P. fragi, L. monocytogenes, B. subtilis and Enterococcus attached in biofilm to stainless steel more resistant to disinfectants and sanitizers than unattached cells.
- Portion of attached cells may survive a heat treatment of 70 °C for 5 min.

Microbial Attachments and Biofilms





- Age of biofilm affects the resistance of microorganisms to sanitizers
- Na-hypochlorite and Quats effective against a
 24 hr. biofilm of L.m. on food contact surfaces
- Resistance to chlorine increase with biofilm age

"... It is important that sufficient emphasis be placed on correct cleaning and sanitizing procedures in food processing systems."

Microbial attachment and Biofilm formation IFT Scientific Status Summary, July, 1994.

Environmental Applications

- Indicator and Specific Pathogen testing
- Listeria spp. or Listeria-like organisms, E. coli,
 E. coli 0157:H7
- ▶ Petrifilm™ Environmental Listeria Plate
- > P/A, semi-quantitative or quantitative results

Environmental monitoring for Listeria

Duiry, Food and Environmental Sanitation, Vol. 19, No. 8, Pages 551-562 (applight) MMFS, £200 Jaron Ive., Soits 2009, Des Maines, W. 50322

Guidelines to Prevent Post-Processing Contamination from Listeria monocytogenes

R. Bruce Tompkin, Virginia N. Scott, 2* Dane T. Bernard, 2 William H. Sveum, 3 and Kathy Sullivan Gombas⁴

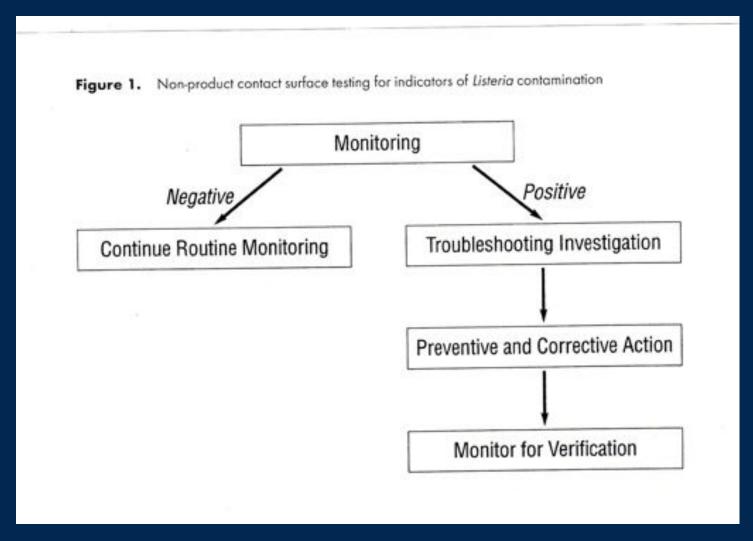
SUMMARY

Extensive efforts to control Listeria monocytogenes can reduce the frequency and level (CFU/g or cm²) of contamination, but it is not possible, given currently available technology, to eradicate it from the processing environment or totally eliminate the potential for contamination of finished products. Because of the serious nature of listeriosis in the susceptible population, industry must take stringent measures to control L. monocytogenes

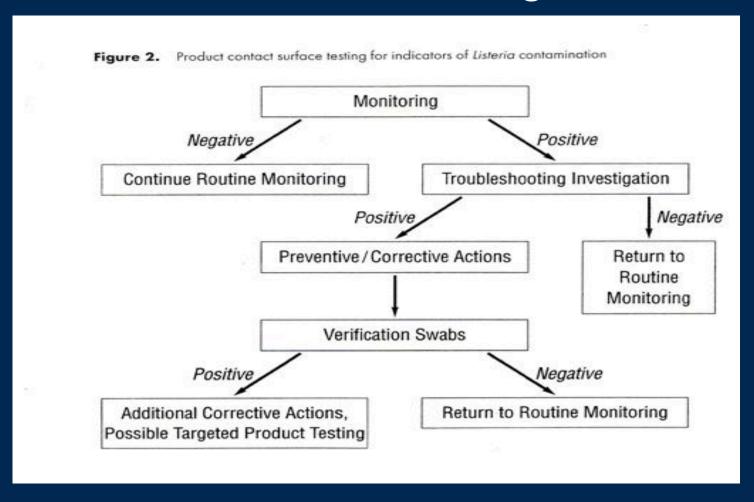
INTRODUCTION

This document is intended to apply to refrigerated, ready-to-eat (RTE) foods that support the growth of Listeria monocytogenes, although the guidelines may be applied to other products to minimize contamination with L monocytogenes. However, not all the guidelines listed below apply in all situations. The controls for L monocytogenes will be product, process and plant specific; therefore, these

Environmental monitoring for Listeria



Environmental monitoring for Listeria

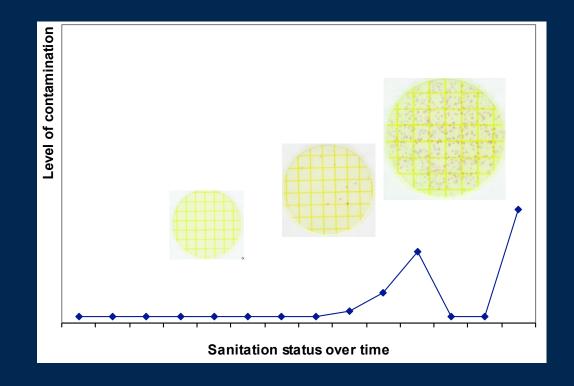


Environmental Applications

> P/A, semi-quantitative or quantitative results







Application of ATP Swabs and ELP plates





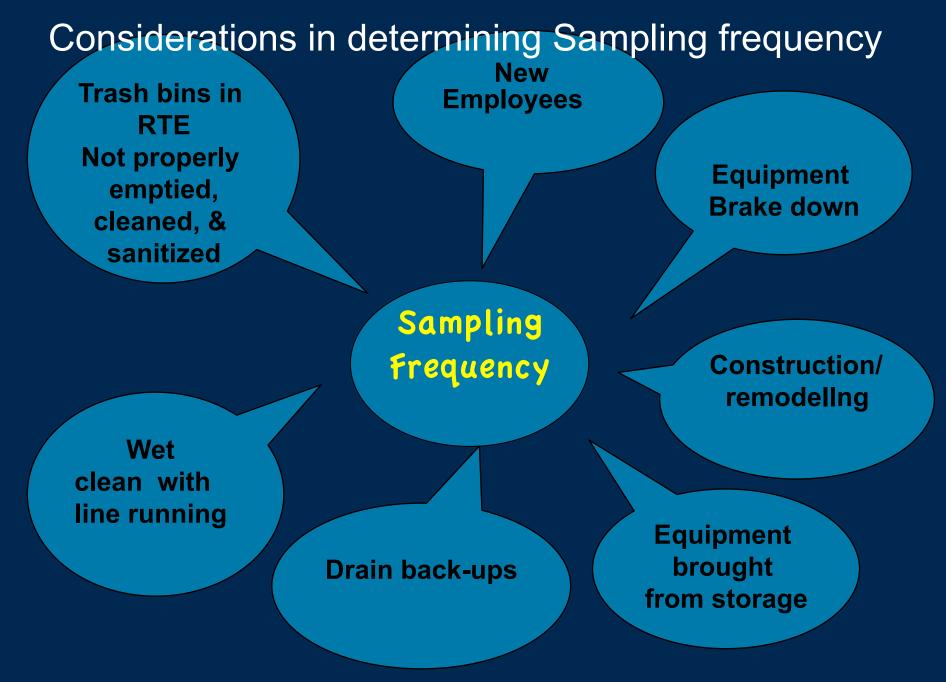


Environmental Monitoring

- Program developed using sponges/swabs to assess microbiological environment of plant
- Used to assess if sanitation procedures are effective
- ➤ Identifies key areas to monitor for presence of bacteria
- ➤ Sanitation measures can be modified to avoid potential product contamination

Consideration in Environmental Monitoring

- >EM program includes several aspects
 - Sampling frequency
 - Pre-Op vs. Operation samples
 - Sampling sites
 - Sampling techniques
 - Sample area size



Sample Size

- >Ideal sample size
 - >40 Square inches of area with sponge/swab
 - ➤ Use horizontal and vertical motion
 - ➤ Pressure should not cause the sponge to crumble
 - ➤ Smaller sample area is acceptable for area that is not easily sampled

Sampling sites

- ➤ Non-product and product contact surfaces are selected.
- ➤Zone Concept

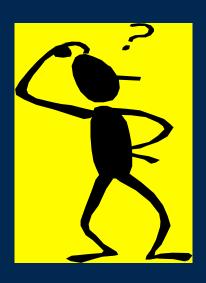
Summary and Conclusions

- Main objective of cleaning and sanitation is to control Microorganisms
- Effective sanitation programs include monitoring sanitation efficacy
- Monitoring may be done visually or by checking cleaning conditions but microbiological testing can confirm cleaning and sanitation efficacy
- Several methods for microbiological monitoring are available
- ATP hygiene monitoring and zone testing are popular option for hygiene monitoring
- ATP bioluminescence based methods gives Total cleanliness of surfaces (microorganisms + food residue) but readings may not correlate with SPC
- > Protein Swabs give "relative" data, Hard to quantify

Acknowledgements..

- > IAFP and the Dairy Sanitaion PDG
- Various companies marketing microbiological testing equipment and supplies. Mention of a brand name does not necessarily imply endorsement.
- UW River Falls

Any Questions ???



Thank You!!

