# biogrowth & bioinactivation - making predictive microbiology a bit easier

### **IAFP Software Fair Series**

Alberto Garre, postdoc at Wageningen University







We know our RTE product is safe... but One day, is was on the sun for one hour

## What should we do?





### **Predictive microbiology**

Part of food science that develops and validates mathematical models to predict the microbial response within the food chain



## Predictive microbiology is mature now

 ${\sim}100~\text{or}$  50 years old

Main principles defined

Established experimental methodologies

Sheared modeling approaches

Included in some regulations





## (Some) applications of predictive micro

Process design

Shelf life estimation

HACCP plans

Food Safety Objectives

Quantitative Microbial Risk Assessment Education





### The usual modelling approach

**Primary model**: relationship between the microbial count and the elapsed time **Secondary model**: relationship between the kinetic parameters and the environment (temperature, pH...)



## Model fitting is central in predictive microbiology

Parameters cannot be known

They are estimated from data

Always question validity





## Issue: predictive micro cannot be done by hand



Methodology well established... but complex

Fitting of non-linear models can be complicated (identifiability).

Dynamic growth requires models in differential equations.



## Software can bridge this gap

### During the last years several applications to make life easier

### Simplify and facilitate the model process

### Cover practically every step of QMRA/shelf life estimation



Food Microbiology Volume 45, Part B, February 2015, Pages 290-299



Software for predictive microbiology and risk assessment: A description and comparison of tools presented at the ICPMF8 Software Fair

Fanny Tenenhaus-Aziza ª ⊠, Mariem Ellouze <sup>b</sup> A ⊠

New software solutions for microbiological food safety assessment and management

Current Opinion in Food Science

Volume 44, April 2022, 100814

Arícia Possas 🖾 , Antonio Valero 🖾 , Fernando Pérez-Rodríguez



## biogrowth & bioinactivation

Main goal: facilitate predictive microbiology

- Model fitting & calculation of predictions
- Isothermal & dynamic conditions
- Deterministic & stochastic
- Model comparison





### Who may use these tools?



## An architecture to make everyone happy

### An R package

- Core functions for predictive microbiology
- Integration in other workflows, automation...
- Flexibility





### An online application

- User-friendly interface to the R package
- Additional functions for model interpretation
- Ease of use



## Transparency as a principle

print.DynamicGrowth <- function(x,) {
cat("Growth prediction under dynamic conditions\n\n")
env <- names(v\$env conditions)
cat(paste("Environmental factors included:", paste(env. collapse = ", "), "\n\n"))
cat("Parameters of the primary model:\n")
<pre>print(unlist(x\$primary_pars))</pre>
cat("\n")
for /i in 1:longth/v@coo modele\) [
<pre>cat(paste("Secondary model for ", names(x\$sec models)[i], ":\n", sep = ""))</pre>
<pre>print(unlist(x\$sec_models[[i]]))</pre>
cat("\n")
}

#### **Detailed manuals**

This module includes a variety of primary inactivation models commonly used in predictive microbiology. The user can select among them using  $\mathbb{O}$ . Then, the values of the model parameters must be defined using the numeric inputs  $\mathbb{Q}$ . Note that the contents of this box are updated according to the inactivation model selected. For details about the models implemented, please look at Appendix A.



#### **Implementation details**

#### (Adapted) Full Ratkowsky model

The full Ratkowsky model (Ratkowsky et al., 1983) is an extension of the square-root model by Ratkowsky (Ratkowsky et al., 1982) that accounts for the decline of the growth rate for temperatures higher than the optimal one. It is described by the following equation

$$\sqrt{\mu} = b(X - X_{min})(1 - e^{c(X - X_{max})})$$

where X is the value of the environmental factor,  $X_{min}$  is the theoretical minimum value for growth,

 $X_{max}$  is the theoretical maximum value for growth, b is a parameter that defines the slope of the curve for suboptimal temperatures and c defines the slope for superoptimal temperatures.

Due to structural identifiability and compatibility with the other models, to use it as a gamma factor, this model must be modified so it is defined within [0, 1]. This can be done by defining a function such as

 $\gamma_{Ratkowsky} = \left(\frac{\sqrt{\mu(X)}}{\sqrt{\mu}}\right)^2$ 

#### **Code examples**

Because the temperature during the simulation equals the cardinal parameter  $X_{opt}$ , the predicted population size is identical to the one calculated using predict\_isothermal\_growth for the Baranyi model when  $\mu = \mu_{opt}$  and  $\lambda = \frac{\ln(1+1/Q_0)}{\mu_{opt}}$ .

lambda <- Q0\_to\_lambda(q0, mu\_opt)</pre>

my\_model <- "Baranyi"
my pars <- list(logN0 = 2, logNmax = 8, mu = mu opt, lambda = lambda)</pre>

static\_prediction <- predict\_isothermal\_growth(my\_model, my\_times, my\_pars)</pre>

plot(static\_prediction) +





## Features of biogrowth

#### Predictions

- Isothermal conditions
- Parameter uncertainty
- Dynamic conditions
- Any number of factors
- Several models

#### Other

- Visualization
- Statistical summaries
- Goodness of fit
- Results export

#### Model fitting

- Primary models
- Dynamic experiments
- Global fitting
- Cardinal parameters
- Fixing any parameter
- Any number of factors
- Several models

### Isothermal growth





## Primary models in *biogrowth*

#### Baranyi

$$\log N(t) = \log N_{max} + \log \frac{1 + \exp(\ln(10)\mu(t-\lambda)) - \exp-\ln(10)\mu\lambda}{\exp(\ln(10)\mu(t-\lambda)) - \exp(-\ln(10)\mu\lambda) + 10^{\log N_{max} - \log N_0}} \qquad \log N(t) = \begin{cases} \log N_0 & \text{if } t < \lambda \\ \log N_{max} & \text{if } t > t_{stat} \\ \log N_0 + \mu(t-\lambda) & \text{otherwise} \end{cases}$$

**Trilineal** 

#### modified Gompertz

$$\log N(t) = \log N_0 + C\left(\exp\left(-\exp\left(\frac{e \cdot \mu}{C}(\lambda - t) + 1\right)\right)\right)$$

#### Logistic

$$\log N(t) = \log N_0 + \frac{C}{1 + \exp\left(\frac{4\mu}{C}(\lambda - t) + 2\right)}$$

#### **Richards**

$$\log N(t) = \log N_0 + C \left[ 1 + \nu \cdot \exp\left(1 + \nu + \frac{\mu}{C}(1 + \nu)^{1 + 1/\nu}(\lambda - t)\right) \right]^{-1/\nu}$$



### Secondary modelling - gamma concept

#### **Hypotheses:**

- each suboptimal factor (pH, aw, T...) reduces the growth rate
- each effect is independent

$$\mu(t) = \mu_{opt} \cdot \gamma_1(X_1(t)) \cdot \ldots \cdot \gamma_n(X_n(t)); i \in 1, \dots, k$$



#### **Models included**

- Cardinal parameter model
- Full Ratkowsky model
- Zwietering-type models

## The biogrowth R package

#### Available from CRAN

#### install.packages("biogrowth")

#### Extensively documented





#### biogrowth: Modelling of Population Growth

Modelling of population growth under static and dynamic environmental conditions. Includes functions for model fitting and making prediction under isothermal and dynamic conditions. The methods (algorithms & models) are based on predictive microbiology (See Perez-Rodriguez and Valero (2012, ISBN:978-1-4614-5519-6)).

Depends: R (≥ 2.10)	
Imports:         deSolve (≥ 1.28), tibble (≥ 3.0.3), dplyr (≥ 0.8.5), FME (≥ 1.3.6), MASS (≥ 7.3), rlan           0.3.4), ggplot2 (≥ 3.3.2), cowplot (≥ 1.0.0), lamW (≥ 1.3.0), tidyr (≥ 1.0.2), formul	ng (≥ 0.4.7), <u>purrr</u> (≥ l <u>a.tools</u> (≥ 1.7.1)
Suggests: <u>knitr, rmarkdown, tidyverse</u> (≥ 1.3.0)	
Published: 2021-01-27	
Author: 🛛 🛛 Alberto Garre 👩 [aut, cre], Jeroen Koomen [aut], Heidy den Besten [aut], Mar	cel Zwietering [aut]
Maintainer: Alberto Garre <garre.alberto at="" gmail.com=""></garre.alberto>	
License: <u>GPL-3</u>	
NeedsCompilation: no	
Materials: <u>README NEWS</u>	
CRAN checks: biogrowth results	
Downloads:	
Reference manual: <u>biogrowth.pdf</u>	
Vignettes: Publication ready figures with biogrowth	
Modelling population growth with biogrowth	
Using dynamic models for static environmental conditions	
Package source: <u>biogrowth_0.2.0.tar.gz</u>	
Windows binaries: r-devel: <u>biogrowth_0.2.0.zip</u> , r-release: <u>biogrowth_0.2.0.zip</u> , r-oldrel: <u>biogrowth</u>	_0.2.0.zip
macOS binaries: r-release (arm64): <u>biogrowth_0.2.0.tgz</u> , r-release (x86_64): <u>biogrowth_0.2.0.tgz</u> ,	r-oldrel:
<u>biogrowth_0.2.0.tgz</u>	
Old sources: <u>biogrowth archive</u>	
Linking:	

Please use the canonical form <u>https://CRAN.R-project.org/package=biogrowth</u> to link to this page.

### Just a few lines

```
## We will use the multiple experiments data set
 4
    data("multiple experiments")
 5
 6
    ## For each environmental factor, we need to defined a model
 8
    sec names <- c(temperature = "CPM", pH = "CPM")</pre>
 9
10
    ## Any model parameter can be fixed
11
12
    known <- list(Nmax = 1e8, NO = 1e0, OO = 1e-3,
13
                  temperature n = 2, temperature xmin = 20, temperature xmax = 35,
14
                  pH n = 2, pH xmin = 5.5, pH xmax = 7.5, pH xopt = 6.5)
15
16
   ## The rest require starting values for model fitting
17
18
   start <- list(mu opt = .8, temperature xopt = 30)</pre>
19
20
21
    ## We can now call the fitting function
22
    global fit <- fit multiple growth(start, multiple experiments, known, sec names)
23
24
    ## Parameter estimates can be retrieved with summary
25
26
    summary(global_fit)
27
28
    ## Any single environmental factor can be added to the plot using add factor
29
30
    plot(global_fit, add_factor = "temperature")
31
22
```



Parameters:			
	Estimate Std.	Error t value P	r(> t )
mu_opt	0.54196 0	.01222 44.35	<2e-16 ***
temperature_xopt	30.62396 0	.18728 163.52	<2e-16 ***
Signif. codes:	0 '***' 0.001	'**' 0.01 '*' O.	05'.'0.1''1
Residual standar	d error: 0.428	2 on 48 degrees	of freedom
Parameter correl	ation:		
	mu_opt tempera	ature_xopt	
mu_opt	1.0000	0.8837	
temperature_xopt	0.8837	1.0000	
1		Bol	
	••••	u0 1	• • •



## The online version of *biogrowth*

https://foodmicrowur.shinyapps.io/biogrowth/

#### PC

WAGENINGEN

VIVERSITY & RESEARCH









### Open code



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<> (	Code 💿 Issues	្រាំ Pull requests	<ul> <li>Actions</li> </ul>	III Projects	🖽 Wiki	
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	R	Updated the labels of plot.	FitSecond	10 days ago	🛄 Readr	
	data	Included two new datasets		21 days ago		
	man	Included two new datasets		21 days ago	Releases	3
	vignettes	Updated the parameters of	f the vign	10 days ago	No releases published Create a new release	
	.Rbuildignore	Improved the README file		7 months ago		
	.gitignore	Small changes for CRAN		8 months ago	Package	s
	DESCRIPTION	Added times to predict.Fit	)ynamicG	4 months ago	No packages published	
	NAMESPACE	Implemented formula inter	face for pr	5 months ago	Publish your	first package
	NEWS.md	Updated the NEWS		10 days ago	Contribu	
	README.Rmd	Updated the README file		4 months ago	Contribu	
	README.md	Edited README.md		4 months ago		garre Alberto
	cran-comments	Prepared for CRAN		7 months ago	Jero	<b>ben-K</b> Jeroen

https://g	ithub.com/albga rowth_web	watch + 1	Star 1 ए Fork	ve
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ilbgarre Added	a help page to the static predi	days ago 🕚 35	or topics provided.	
🖿 man	Added a help page to the static pred	11 days ago	🛱 Readme	
B README.Rmd	Improved the README file	7 months ago		
README.md	Improved the README file	7 months ago	Releases No releases published Create a new release	
🗋 example_cardi	Fitting cardinal parameters	9 months ago		
🗋 example_dyna	. Dynamic prediction	9 months ago		
🗋 example_dyna	Added a few changes to make every	8 months ago	Packages	
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🗋 example_dyna	Added a few changes to make every	8 months ago		
🗋 example_globa	. I had forgotten to commit the examp	7 months ago	Languages	
example_globa	. I had forgotten to commit the examp	7 months ago		
example_static	Added a few changes to make every	8 months ago	• <b>R</b> 100.0%	
my_body.R	Added a help page to the static pred	11 days ago		
server.R	Added a help page to the static pred	11 days ago		

## Live demonstration

#### https://foodmicrowur.shinyapps.io/biogrowth/





- Fitting primary growth models
- Fitting cardinal growth models (temperature & pH)
- Validation under dynamic conditions
- Fitting dynamic growth models







## Features of *bioinactivation*

### Predictions

- Isothermal conditions
- Dynamic conditions
- Parameter uncertainty
- Several models

#### Other

- Visualization
- Statistical summaries
- Goodness of fit
- Results export

### Model fitting

- Primary models
- Two-step fitting
- One-step fitting
- Dynamic models
- Fixing any model parameter
- Several models

#### Isothermal models included in *bioinactivation* **Bigelow** Geeraerd $\log N = \log N_{res} + \log 10(((10^{\log N0 - \log Nres} - 1) \cdot e^{k \cdot SL})/(e^{k \cdot t} + e^{k \cdot SL} - 1) + 1)$ $\log N = \log N_0 - t/D$ Peleg 2 pop. Weibull $\log N = \log N_0 - b \cdot t^n$ $N = N0/(1+10^{\alpha})[10^{-(t/\delta 1)^{p_1} + \alpha} + 10^{-(t/\delta 2)^{p_2}}]$ Mafart Metselaar $\log N = \log N_0 - (t/\delta)^p$

#### Trilinear model

 $\begin{array}{l} logN = logN_0; \, t < SL \\ logN = logN_{res}; \, t > t_{res} \\ logN = logN_0 - (t-SL)/D; \, otherwise \end{array}$ 



## Dynamic models included in *bioinactivation* Bigelow Mafart

$$\frac{d\log_{10}S}{dt} = -p\left(\frac{1}{\delta(T)}\right)^p t^{p-1}$$

$$\log_{10} D(T) = \log_{10} D(T_{ref}) - \frac{T - T_{ref}}{z}$$

$$\log_{10} D(T) = \log_{10} D(T_{ref}) - \frac{T - T_{ref}}{z}$$

#### Geeraerd

 $\frac{d\log_{10}N}{dt} = -\frac{1}{D(T)}$ 

$$\frac{dN}{dt} = -\frac{1}{1+C_c} k_{max}(T) N \left(1 - \frac{N_{res}}{N}\right)$$
$$k_{max}(T) = k_{max}(T_{ref}) 10^{-(T-T_{ref})/z}$$

$$\frac{d\log_{10}S}{dt} = -b(T) n \left(\frac{-\log_{10}S}{b(T)}\right)^{\frac{n-1}{n}}$$

$$b(T) = \ln \left(1 + e^{k_b(T - T_c)}\right)$$



## The bioinactivation R package

#### Available from CRAN

install.packages("bioinactivation")

#### Extensively documented

https://github.com/albgarre/bioinactivation

bioinactivation: Mathematical Modelling of (Dynamic) Microbial Inactivation

Functions for modelling microbial inactivation under isothermal or dynamic conditions. The calculations are based on several mathematical models broadly used by the scientific community and industry. Functions enable to make predictions for cases where the kinetic parameters are known. It also implements functions for parameter estimation for isothermal and dynamic conditions. The model fitting capabilities include an Adaptive Monte Carlo method for a Bayesian approach to parameter estimation.

Version:	1.2.3
Imports:	$ \begin{array}{l} \underline{dplyr} (\geq 0.4.1), \ \underline{deSolve} (\geq 1.11), \ \underline{FME} (\geq 1.3.2), \ \underline{lazyeval} (\geq 0.1.10), \\ \underline{gplot2} (\geq 2.0.0), \ \underline{MASS} (\geq 7.3-39), \ graphics (\geq 3.1.3), \ stats (\geq 3.1.3), \ \underline{rlang} (\geq 0.1.2), \ \underline{purrr} (\geq 0.3.2) \end{array} $
Suggests:	<u>knitr</u> (≥ 1.9), <u>testthat</u> (≥ 0.9.1), <u>rmarkdown</u> (≥ 1.12)
Published:	2019-08-01
Author:	Alberto Garre [aut, cre], Pablo S. Fernandez [aut], Jose A. Egea [aut]
Maintainer:	Alberto Garre <garre.alberto at="" gmail.com=""></garre.alberto>
License:	GPL-3
NeedsCompilation	no
Citation:	bioinactivation citation info
Materials:	NEWS
CRAN checks:	bioinactivation results



### Just a few lines

```
1
 2
    data(dynamic inactivation) # The example data set is used.
 3
    simulation model <- "Peleg" # Peleg's model will be used</pre>
 4
 5
 6
    dummy_temp <- data.frame(time = c(0, 1.25, 2.25, 4.6),
 7
                              temperature = c(70, 105, 105, 70) # Dummy temp. profile
 8
 9
    ## Set known parameters and initial points/bounds for unknown ones
10
11
    known_params = c(temp_crit = 100)
12
13
    starting_points <- c(n = 1, k_b = 0.25, N0 = 1e+05)
    upper_bounds <- c(n = 2, k_b = 1, N0 = Inf)
14
    lower_bounds <- c(n = 0, k_b = 0, N0 = 1e4)
15
16
17
    dynamic_fit <- fit_dynamic_inactivation(dynamic_inactivation, simulation_model,</pre>
18
                                              dummy_temp, starting_points,
19
                                              upper bounds, lower bounds,
20
                                              known_params)
21
22
    plot(dynamic fit)
23
    summary(dynamic_fit)
24
```

```
Parameters:
    Estimate Std. Error t value Pr(>|t|)
n 4.186e-01 7.757e-02 5.397 4.12e-06 ***
k_b 6.596e-01 3.212e-02 20.531 < 2e-16 ***
N0 1.213e+05 3.137e+04 3.865 0.000433 ***
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 0.4328 on 37 degrees of freedom



## The web version



16

#### bioinactivation4

Complete interface to the package Improved & modernized interface Additional editing tools

#### bioinactivation FE

Extended functions One-step fitting Dynamic predictions

#### bioinactivation SE

Illustration of the package Fitting dynamic experiments



### Open code



#### https://github.com/albgarre/higipactivation

albgarre	bgarre / bioinactivation (Public)				
> Code	⊙ Issues ൂn Pull requests ⊙	Actions 🖽 Projects 🖽 Wiki 🔅	) Security 🖂 Insights 🛞 Settings		
	master - 1 branch 🚫 1 tag		Go to file Add file - Code -		
	albgarre Got rid of all the mutate		b6746c6 on Jul 5, 2021 🕚 124 commits		
	data	Updated training set documentation.			
	inst				
	man				
	tests				
	vignettes				
۵	.gitignore	Included graphics in ggplot2.			
۵	DESCRIPTION				
۵	NAMESPACE				
۵	NEWS.md				
۵	README.Rmd	Extended README file.			
۵	README.md	Update README			
۵	cran-comments.md	Prepared for CRAN re-submission			

README.md

bioinactivation: a package for modelling microbial inactivation in R.

The bioinactivation package implements several function for the modelization of microbial inactivation in R. It includes:

htt	https://aithub.com/albaarre/bioinactivation4				
lbgarr	re / bioinactivation4 Public				
Code	⊙ Issues 🏥 Pull requests	🕞 Actions 🛗 Projects 🛄 Wiki	🕐 Security 🛛 🗠 Insights 🎄 Settings		
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	R				
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C	ງ .gitignore				
C	ງ LICENSE				
C	ງ README.md	Updated the README file	14 days ago		
C	ງ Rplot01.png	Updated application			
C	example_1step.csv	Updated application			
C	example_1step.xlsx	Updated application			
C	) server.R	Implemented the 2 step module			
C	ງ ui.R	Added a link to the manual	14 days ago		

#### README.md

ß

#### bioinactivation4

The bioinactivation package for R includes several function for building and applying microbial inactivation models (Garre et al., 2017). This package is available on CRAN, making it easy to install in any computer with R. The package is Open Source, with the source code available on GitHub:

## Live demonstration

https://foodlab-upct.shinyapps.io/bioinactivation4/





- Fitting primary inactivation models
- Fitting secondary models: one & two-step approach
- Validation under dynamic conditions
- Fitting dynamic inactivation models



## Additional reading



Food Research International Volume 93, March 2017, Pages 66-74



Bioinactivation: Software for modelling dynamic microbial inactivation

Alberto Garre ª, Pablo S. Fernández ª A $\boxtimes$ , Roland Lindqvist  $^{\rm b}$ , Jose A. Egea  $^{\rm c}$ 



Food Research International Volume 112, October 2018, Pages 353-360



Bioinactivation FE: A free web application for modelling isothermal and dynamic microbial inactivation

Alberto Garre ª, Marta Clemente-Carazo ª, Pablo S. Fernández ª 🖾, Roland Lindqvist <sup>b</sup>, Jose A. Egea <sup>c</sup> 🖄 🖾



# Roadmap 2022 2023

**biogrowth package** update

#### bionactivation4

release



**biogrowth web** optimize new features release

**bionactivation** modernize

## Conclusions

- bioinactivation and biogrowth try to make life easier
- They can ease focusing on the most relevant parts of a study
- They are free, open and transparent
- Users decide software quality
- Your feedback is essential for improving the tools



## Acknowledgements



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- Xuchuan Ma, George Pampoukis (WUR)

#### alberto.garreperez@wur.nl









## backup slides







## Fitting primary models







## Isothermal fitting

