

A Compilation of Histamine-Forming Bacteria Associated with Foods

John DeBeer^{1*} and Jon W. Bell²

¹Retired, Encinitas, CA, USA

²National Seafood Inspection Laboratory, National Marine Fisheries Service, National Oceanic and Atmospheric Administration, 3209 Frederic St., Pascagoula, MS, 39567 USA

SUMMARY

The objective of this article is to document the names of histamine-forming bacteria (HFB) found in foods and available from the scientific literature. This extensive compilation found 148 unique species of HFB identified, representing 57 genera, 25 families, 13 orders, 6 classes, and 3 phyla. A synonymous species name was recorded in many cases, so both species names have been recorded in this article. Based on the findings, there were 78 species of Gram-negative and 70 species of Gram-positive HFB, with 18 spore-forming species. The food matrix, or type of food from which the bacteria were identified, also is provided, and the most common food matrix identified in this listing was fish based. The genera with the most HFB species were *Bacillus* (14), *Lactobacillus* (11), and *Staphylococcus* (8). The families with the most genera were *Lactobacillaceae* (31), *Enterobacteriaceae* (29), and *Vibrionaceae* (17). A list of published HFB, publication authors, and corresponding document DOI numbers is included. This compilation with DOI numbers should be useful to other researchers searching for HFB with certain characteristics.

INTRODUCTION

The objective of this article is to document histamine-forming bacteria (HFB) that have been cited in published literature and are present in human foods. The effort and study objectives resulted from various investigations over the years that the authors have been involved in, looking for HFB with particular characteristics. This initial effort found no single database of HFB and characteristics in human food, so a comprehensive list of HFB was compiled.

Histamine can be formed in certain bacteria by the enzyme histidine decarboxylase (HDC) if free molecules of the amino acid L-histidine are present. HDC is uniquely able to remove a carbon dioxide molecule from the L-histidine molecule to form histamine (51), and either of two pathways are used, depending on whether the bacteria are Gram-positive or Gram-negative. The Gram-positive bacteria use a pyruvoyl residue that is covalently bound at

the active site, whereas pyridoxal phosphate-dependent enzymes are used in Gram-negative bacteria (25). The bacteria gain a proton of energy and pH control from this decarboxylase reaction (34, 35). Previous authors provide a clear explanation and schematic of the cellular process to produce histamine in the bacteria (17, 30, 35). This process includes the proton motive force generated by the decarboxylation and electrogenic antiporter mechanism of transporting the L-histidine molecule into the bacterial cell, the ensuing decarboxylation reaction, and the histamine molecule that is produced and then transported or exported out of the cell. A generalized scheme is presented by Molenaar et al. (34).

The histidine → histamine reaction requires free L-histidine molecules to be available in the food matrix for the HFB to process. This histidine molecule is provided by either the naturally occurring free L-histidine in muscle tissues or the food matrix. The free L-histidine also is available during fermentation when proteins are broken down (proteolysis) into their free amino acids (41). For example, free L-histidine is naturally available in the muscle tissue of scombroid and other fast-moving fish. Large amounts of free L-histidine in these fish act as a buffer during fight-or-flight muscle movement (9). However, other sources of free amino acids, such as histidine, are generated during the fermentation of foods such as kimchi, miso, cheese, sausage, wine, beer, and fermented fishery products. In virtually all foods containing proteins and experiencing the conditions that enable microbial fermentation activity, histamine formation can be expected if HFB are present. The total amount of the histamine formed strongly depends on the nature of the food, types of bacteria present, and incubation temperature (30).

Histamine is the only biogenic amine that is regulated for food safety (60), and it is only regulated for fish, primarily scombroid fish (19). The allowable sets of limits for histamine content in fish varies in different countries. For example, the levels of histamine allowed in fish in the United States differ from those in the European Union (12).

*Author for correspondence: Phone: +1 760.670.6141; E-mail: jdebeer2005@gmail.com

TABLE 1. Species or genera (unique) that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability

Gram-positive or Gram-negative	Spores	Environment	Count	Subtotal	Total
Positive	Endospore	Aerobic	2		
		Anaerobic	2		
		Facultative anaerobic	14	18	
	No	Aerobic	2		
		Aerotolerant	11		
		Anaerobic	3		
		Facultative anaerobic	32		
		Microaerophile	2		
		Unknown	2	52	70
Negative	No	Aerobic	15		
		Facultative aerobic	3		
		Facultative anaerobic	60	78	78
Total unique bacterial names			148		

A list of histamine-secreting bacteria in the human gut microbiome has been developed based on genomic analyses (35). Although the lists may overlap, this article focuses on HFB in human foods.

METHODS

An extensive literature search of the electronic media using Google Scholar and other search engines was conducted using terms such as HFB, tuna, scombroid fish poisoning, cheese, sausage, fermented foods, sauerkraut, spinach, fermented fish, and wine. The HFB species were recorded and then identified as to genus, family, order, class, and phylum. Characteristics such as whether the species is Gram-positive or Gram-negative, whether it is a spore former, and its preferred level of oxygen in the environment also were noted. The data were recorded in Microsoft Excel tables and summarized using pivot tables. The species, genera, and some characteristics were verified with the website BacDive (2).

Not every published paper that mentions HFB can be cited or mentioned here; however, when a species new to the list being developed was found, the entire list of HFB from that paper was included in the dataset, along with all characteristics of that species or group. Some HFB activity in foods was only identified to the genus or family level, and genera or families were included in the master list as well. Several HFB species were listed in only one source and were not identified in BacDive (2), so some characteristics were labeled as unknown.

RESULTS

In total, 182 species of bacteria, genus, or family names were identified as HFB present in human foods and are listed in [Appendix A](#). Bacterial names are updated periodically, and current correct names and synonyms have been previously documented and collated in BacDive (2). There are four types of listing for the HFB in this article: the correct species names and synonyms; synonyms only; one unique genus; and one unique family. The unique species, genera, and families are identified in [Appendix A](#), as well as characteristics such as Gram-positive or Gram-negative, oxygen requirements, and the ability to form spores. Of the 148 unique names of species and/or genera, there were 70 Gram-positive and 78 Gram-negative species ([Table 1](#)). The Gram-positive group contained 18 species that can form spores under certain conditions. The 148 HFB species were part of 57 genera, 25 families, 13 orders, 6 classes, and just 3 phyla ([Table 2](#)). For comparison it is estimated that there are at least 65 known bacterial phyla (47).

It's important to understand the environment (oxygen requirements) for these HFB. For example, *Tetragenococcus* spp. are capable of growing in anaerobic environments, like unretorted canned anchovies (4) (DeBeer, J., personal communication). Many *Bacillus* spp. can form spores that withstand harsh dry environments, like dried fishmeal (23). This is not a complete list, but the compilation hopefully can help future researchers understand why they suddenly find an unfamiliar species in food (46).

TABLE 2. Taxonomic groupings of histamine-forming bacteria

Taxonomic grouping	Count
Species	148
Genera	57
Families	25
Orders	13
Classes	6
Phyla	3

TABLE 3. Environment, food types, and food matrices for histamine-forming bacteria activity

Environment	Food type	Food matrix	Count	Subtotal	Total
Land based	Fermented food	Fermented food (cheese, meat, & wine)	20		
		Fermented foods	15	35	
	Fermented food, dairy	Cheese	18		
		Cheese, goat	2		
		Fermented food, cheese	8		
		Fermented food, cheese & other foods	1		
		Fermented food, cheese biofilms	1		
		Fermented food, Italian cheese	1		
		Fermented food, semihard Tilsit (Swiss) cheese	2		
		Fermented food, dairy	1	34	
	Fermented food, vegetable	Vegetable, kimchi	1		
		Vegetable, miso	5		
		Vegetable, mustard pickle	3		
		Vegetable, soy sauce	1		
		Vegetable, spinach	5		
		Vegetable, sugar beets	2	17	86
Marine	Seafood	Fermented fish products	2		
		Fermented Malaysian fish sausage	3		
		Fermented seafood & vegetable, salted	2		
		Fermented seafood, salted canned anchovies	5		
		Seafood	4		
		Seafood, <i>Auxis thazard</i>	4		
		Seafood, cod	2		
		Seafood, dried flying fish	1		
		Seafood, fish	18		
		Seafood, fish paste	22		

Table continued on the next page.

TABLE 3. Environment, food types, and food matrices for histamine-forming bacteria activity (cont.)

Environment	Food type	Food matrix	Count	Subtotal	Total
Marine	Seafood	Seafood, Indian mackerel	21		
		Seafood, longtail & eastern little tuna	14		
		Seafood, multiple fish species	67		
		Seafood, Pacific mackerel	11		
		Seafood, sailfish	14		
		Seafood, sailfish & milkfish	1		
		Seafood, salted dried fish	7		
		Seafood, sardines	6		
		Seafood, scombroid	10		
		Seafood, shrimp, marine, brackish	25		
		Seafood, swordfish	2		
		Seafood, tuna sausage	8		
		Seafood, tuna waste	7		
		Seafood, various sources	25		
		Seafood, yellowfin tuna	10	291	
	Ocean water	Water samples, Gulf of Mexico	16	16	307
Overall total				393	

This article does not list, quantify, or compare the histamine formation of the HFB, because this ability is not often detailed in the published literature. In addition, individual species are not identified as psychrophilic, mesophilic, or thermophilic, because bacterial serovars of some species can assume more than one of these three conditions. For example, *Weissella paramesenteroides*, a known HFB, is identified as being psychrophilic, mesophilic, or thermophilic, depending on the strain involved (2).

All of these HFB include a notation that the species name was correct for mid-2024 or identified in the literature as a synonym name ([Appendix A](#)). Multiple species were found that were identified with different names (synonyms) by different authors at different periods. These HFB are listed as synonyms in [Appendix A](#). The number of species with synonyms identified in a search of the available literature was 35. [Appendix B](#) lists the HFB with the authors and DOI numbers of the source documentation.

HFB were identified in a multitude of food matrices. [Table 3](#) is divided into 3 sections: environment, food types, and food matrices. The food matrices within the seafood type had by far the most citations of HFB, with 291 mentions by all bacterial species or genera recorded. Fermented foods, including fermented seafood, were cited

in association with 98 bacterial species or genera. Many citations are expected to be duplicates, but an order of magnitude is provided for the sources of HFB within food matrices and/or types. The phyla, classes, orders, families, and genera of the unique HFB are listed in [Table 4](#).

DISCUSSION

About 30,000 formally named bacterial species have been cultured, but there are likely a million to a billion unnamed species (15). Curtis et al. (11) estimate that there are 2 million bacterial taxa in the sea. As can be readily seen by what has been provided in this compilation, the number of HFB, with fewer than 150 named species, is a tiny percentage of the total number of bacterial species.

In this compilation of HFB, the genera with the highest numbers of unique species were *Bacillus* (14), *Lactobacillus* (11), and *Staphylococcus* (8). The families with the highest numbers of genera were *Lactobacillaceae* (31), *Enterobacteriaceae* (29), and *Vibrionaceae* (17). The orders with the most reported HFB species were Enterobacterales (44), Lactobacillales (39), Bacillales (27) and Vibrionales (17).

In conclusion, the authors hope this compilation of HFB is useful to future researchers when they are studying an unknown source of histamine. Both authors have had occasions in the past in which they could have used this list.

TABLE 4. Phylum, class, order, family, and genus of all unique histamine-forming bacteria

Phylum	Class	Order	Family	Genus	Total
Actinomycetota	Actinomycetia	Micrococcales	<i>Brevibacteriaceae</i>	<i>Brevibacterium</i>	1
			<i>Micrococcaceae</i>	<i>Micrococcus</i>	1
Bacillota	Bacilli	Bacillales	<i>Bacillaceae</i>	<i>Bacillus</i>	14
				<i>Lysinibacillus</i>	1
			<i>Listeriaceae</i>	<i>Listeria</i>	1
			<i>Paenibacillaceae</i>	<i>Brevibacillus</i>	1
			<i>Staphylococcaceae</i>	<i>Staphylococcus</i>	10
		Lactobacillales	<i>Enterococcaceae</i>	<i>Enterococcus</i>	2
				<i>Tetragenococcus</i>	2
			<i>Lactobacillaceae</i>	<i>Lactocaseibacillus</i>	2
				<i>Lactiplantibacillus</i>	1
				<i>Lactobacillus</i>	11
				<i>Latilactobacillus</i>	2
				<i>Lentilactobacillus</i>	2
				<i>Lentilactobacillus</i>	2
				<i>Leuconostoc</i>	1
				<i>Levilactobacillus</i>	1
				<i>Limosilactobacillus</i>	2
				<i>Oenococcus</i>	1
				<i>Pediococcus</i>	3
				<i>Weissella</i>	3
			<i>Streptococcaceae</i>	<i>Lactococcus</i>	2
				<i>Streptococcus</i>	2
	Clostridia	Eubacteriales	<i>Lachnospiraceae</i>	<i>Clostridium</i>	2
Pseudomonadota	Alphaproteobacteria	Sphingomonadales	<i>Sphingomonadaceae</i>	<i>Sphingomonas</i>	1
	Betaproteobacteria	Burkholderiales	<i>Alcaligenaceae</i>	<i>Alcaligenes</i>	1
		Neisseriales	<i>Neisseriaceae</i>	<i>Neisseria</i>	1
	Gammaproteobacteria	Aeromonadales	<i>Aeromonadaceae</i>	<i>Aeromonas</i>	3
		Alteromonadales	<i>Shewanellaceae</i>	<i>Shewanella</i>	3
		Enterobacterales	<i>Enterobacteriaceae</i>	<i>Citrobacter</i>	5
				<i>Cronobacter</i>	1
				<i>Enterobacter</i>	6
				<i>Escherichia</i>	2
				<i>Klebsiella</i>	4
				<i>Plesiomonas</i>	1
				<i>Pluralibacter</i>	1
				<i>Proteus</i>	5
				<i>Raoultella</i>	2
				<i>Salmonella</i>	1
				<i>Shigella</i>	1

Table continued on the next page.

TABLE 4. Phylum, class, order, family, and genus of all unique histamine-forming bacteria (cont.)

Phylum	Class	Order	Family	Genus	Total
Pseudomonadota	Gammaproteobacteria	Enterobacterales	Erwiniaceae	Erwinia	1
				Pantoea	2
			Hafniaceae	Edwardsiella	1
				Hafnia	2
			Morganellaceae	Morganella	2
				Providencia	3
			Yersiniaceae	Rahnella	1
				Serratia	3
		Pasteurellales	Pasteurellaceae	Actinobacillus	1
		Pseudomonadales	Moraxellaceae	Acinetobacter	2
				Psychrobacter	1
			Pseudomonadaceae	Pseudomonas	4
		Vibrionales	Vibrionaceae	Listonella	1
				Photobacterium	8
				Vibrio	8
Overall total					148

ACKNOWLEDGMENTS

Each author has worked for U.S. tuna processing companies over the last 30–40 years in various roles, such as procurement, fisheries development, operations, and quality assurance. Each of them has spent many hours in the field working with fishermen and boat owners to try to

prevent histamine growth and maintain seafood freshness in their catches. The authors thank Deborah Mona Baumgartel for her editorial assistance. The authors declare no conflict of interest, financial or otherwise. No external funds were received for this study. A spreadsheet with the listing of HFB names can be obtained from the corresponding author.

REFERENCES

- Ababouch, L., M. E. Afilall, S. Rhafiri, and F. F. Busta. 1991. Identification of histamine-producing bacteria isolated from sardine (*Sardina pilchardus*) stored in ice and at ambient temperature (25°C). *Food Microbiol.* 8:127436. [https://doi.org/10.1016/0740-0020\(91\)90005-M](https://doi.org/10.1016/0740-0020(91)90005-M).
- BacDive. 2023. Global core biodata resource. Available at: <https://bacdive.dsmz.de/>. Accessed 22 March 2024.
- Bang, M. W., C. D. Chung, S. H. Kim, M. B. Chang, S. S. Lee, and S. S. Lee. 2009. Characteristics of histamine forming bacteria from tuna fish waste in Korea. *J. Life Sci.* 19(2):277–283. <https://doi.org/10.5352/JLS.2009.19.2.277>.
- Barbieri, F., C. Montanari, F. Gardini, and G. Tabanelli. 2019. Biogenic amine production by lactic acid bacteria: a review. *Foods* 8(1):17. <https://doi.org/10.3390/foods8010017>.
- Bjornsdottir, K., G. E. Bolton, P. D. McClellan-Green, L. A. Jaykus, and D. P. Green. 2009. Detection of Gram-negative histamine-producing bacteria in fish: a comparative study. *J. Food Prot.* 72(9):1987–1991. <https://doi.org/10.4315/0362-028X-72.9.1987>.
- Bjornsdottir-Butler, K., J. C. Bowers, and R. A. Benner, Jr. 2015. Prevalence and characterization of high histamine-producing bacteria in Gulf of Mexico fish species. *J. Food Prot.* 78(7):1335–1342. <https://doi.org/10.4315/0362-028X.JFP-15-012>.
- Botello-Morte, L., M. Moniente, Y. Gil-Ramírez, R. Virto, D. García-Gonzalo, and R. Pagán. 2022. Identification by means of molecular tools of the microbiota responsible for the formation of histamine accumulated in commercial cheeses in Spain. *Food Control* 133:108595. <https://doi.org/10.1016/j.foodcont.2021.108595>.
- Calles-Enríquez, M., B. H. Eriksen, P. S. Andersen, F. P. Rattray, A. H. Johansen, M. Fernández, V. Ladero, and M. A. Alvarez. 2010. Sequencing and transcriptional analysis of the *Streptococcus thermophilus* histamine biosynthesis gene cluster: factors that affect differential hdcA expression. *Appl. Environ. Microbiol.* 76(18):6231–6238. <https://doi.org/10.1128/AEM.00827-10>.
- Castellini, M. A., and G. N. Somero. 1981. Buffering capacity of vertebrate muscle: correlations with potentials for anaerobic function. *J. Comp. Physiol.* 143:191–198. <https://doi.org/10.1007/BF00797698>.
- Comas-Basté, O., M. L. Latorre-Moratalla, S. Sánchez-Pérez, M. T. Veciana-Nogués, and M. D. C. Vidal-Carou. 2019. Histamine and other biogenic amines in food. From scombroid poisoning to histamine intolerance. *Biogenic Amines* 1. <https://doi.org/10.5772/intechopen.84333>.

11. Curtis, T. P., W. T. Sloan, and J. W. Scannell. 2002. Estimating prokaryotic diversity and its limits. *Proc. Nat. Acad. Sci. USA* 99(16):10494–10499. <https://doi.org/10.1073/pnas.142680199>.
12. DeBeer, J., J. W. Bell, F. Nolte, J. Arcieri, and G. Correa. 2021. Histamine limits by country: a survey and review. *J. Food Prot.* 84(9):1610–1628. <https://doi.org/10.4315/JFP-21-129>.
13. DeBeer, J., F. Nolte, C. W. Lord, and J. Colley. 2021. Tempering large tuna prior to thawing to minimize histamine formation. *Food Prot. Trends* 41(1):36–45. <https://doi.org/10.4315/1541-9576-41.1.36>.
14. Devivilla, S., M. Lekshmi, S. H. Kumar, R. K. Valappil, S. D. Roy, and B. B. Nayak. 2019. Effect of sodium hypochlorite on biofilm-forming ability of histamine-producing bacteria isolated from fish. *J. Food Prot.* 82(8):1417–1422. <https://doi.org/10.4315/0362-028X.JFP-19-101>.
15. Dykhuizen, D. 2005. Species numbers in bacteria. *Proc. Calif. Acad. Sci.* 56(6 Suppl. 1):62.
16. Emborg, J., and P. Dalgaard. 2006. Formation of histamine and biogenic amines in cold-smoked tuna: an investigation of psychrotolerant bacteria from samples implicated in cases of histamine fish poisoning. *J. Food Prot.* 69(4):897–906. <https://doi.org/10.4315/0362-028X-69.4.897>.
17. European Food Safety Authority. 2011. Scientific opinion on risk based control of biogenic amine formation in fermented foods. *EFSA J.* 9:2393. <https://doi.org/10.2903/j.efsa.2011.2393>.
18. Ferrante, M. C., and R. Mercogliano. 2023. Focus on histamine production during cheese manufacture and processing: a review. *Food Chem.* 136046. <https://doi.org/10.1016/j.foodchem.2023.136046>.
19. Food Standards Agency. 2016. Histamine in cheese—Committee on Toxicity. Available at: <https://cot.food.gov.uk/sites/default/files/tox2016-24.pdf>. Accessed 23 April 2024.
20. Frith, A., M. Hayes-Mims, R. Carmichael, and K. Björnsdóttir-Butler. 2023. Effects of environmental and water quality variables on histamine-producing bacteria concentration and species in the northern Gulf of Mexico. *Microbiol. Spectr.* e04720-22. <https://doi.org/10.1128/spectrum.04720-22>.
21. Fröhlich-Wyder, M. T., D. Guggisberg, R. Badertscher, D. Wechsler, A. Wittwer, and S. Irmeler. 2013. The effect of *Lactobacillus buchneri* and *Lactobacillus parabuchneri* on the eye formation of semi-hard cheese. *Int. Dairy J.* 33(2):120–128. <https://doi.org/10.1016/j.idairyj.2013.03.004>.
22. Ginigaddarage, P. H., G. J. Ganegama Arachchi, K. W. S. Ariyawansa, and C. M. Nanayakkara. 2023. Prevalence of histamine forming bacteria in selected nodes of the supply chain of Sri Lankan yellowfin tuna. *J. Nat. Sci. Found.—Sri Lanka* 51(2):215–223. <http://dx.doi.org/10.4038/jnsfsr.v51i2.10819>.
23. Jaw, Y. M., Y. Y. Chen, Y. C. Lee, P. H. Lee, C. M. Jiang, and Y. H. Tsai. 2012. Histamine content and isolation of histamine-forming bacteria in fish meal and fish soluble concentrate. *Fish. Sci.* 78, 155–162.
24. Kim, S. H., K. G. Field, D. S. Chang, C. I. Wei, and H. An. 2001. Identification of bacteria crucial to histamine accumulation in Pacific mackerel during storage. *J. Food Prot.* 64(10):1556–1564. <https://doi.org/10.4315/0362-028X-64.10.1556>.
25. Kimura, B., H. Takahashi, S. Hokimoto, Y. Tanaka, and T. Fujii. 2009. Induction of the histidine decarboxylase genes of *Photobacterium damsela* subsp. *damselae* (formally *P. histaminum*) at low pH. *J. Appl. Microbiol.* 107(2):485–497. <https://doi.org/10.1111/j.1365-2672.2009.04223.x>.
26. Kung, H. F., C. Y. Huang, C. M. Lin, L. H. Liaw, Y. C. Lee, and Y. H. Tsai. 2015. The histamine content of dried flying fish products in Taiwan and the isolation of halotolerant histamine-forming bacteria. *J. Food Drug Anal.* 23(2):335–342. <https://doi.org/10.1016/j.jfda.2014.10.009>.
27. Kung, H. F., Y. H. Lee, D. F. Teng, P. C. Hsieh, C. I. Wei, and Y. H. Tsai. 2006. Histamine formation by histamine-forming bacteria and yeast in mustard pickle products in Taiwan. *Food Chem.* 99(3):579–585. <https://doi.org/10.1016/j.foodchem.2005.08.025>.
28. Kung, H. F., Y. H. Tsai, S. C. Chang, and T. Y. Hong. 2012. Biogenic amine content, histamine-forming bacteria, and adulteration of pork in tuna sausage products. *J. Food Prot.* 75(10):1814–1822. <https://doi.org/10.4315/0362-028X.JFP-12-061>.
29. Kung, H. F., Y. H. Tsai, C. C. Hwang, Y. H. Lee, J. H. Hwang, C. I. Wei, and D. F. Hwang. 2005. Hygienic quality and incidence of histamine-forming *Lactobacillus* species in natural and processed cheese in Taiwan. *J. Food. Drug. Anal.* 13(1):13. <https://doi.org/10.38212/2224-6614.2557>.
30. Landete, J. M., B. De Las Rivas, A. Marcobal, and R. Munoz. 2008. Updated molecular knowledge about histamine biosynthesis by bacteria. *Crit. Rev. Food Sci. Nutr.* 48:697–714. <https://doi.org/10.1080/10408390701639041>.
31. Lavizzari, T., M. Breccia, S. Bover-Cid, M. C. Vidal-Carou, and M. T. Veciana-Nogués. 2010. Histamine, cadaverine, and putrescine produced in vitro by *Enterobacteriaceae* and *Pseudomonadaceae* isolated from spinach. *J. Food Prot.* 73(2):385–389. <https://doi.org/10.4315/0362-028X-73.2.385>.
32. Lim, E. S. 2016. Inhibitory effect of bacteriocin-producing lactic acid bacteria against histamine-forming bacteria isolated from Myeolchi-jeot. *Fish. Aquat. Sci.* 19:1–10. <https://doi.org/10.1186/s41240-016-0040-x>.
33. Ma, J., Y. Nie, L. Zhang, and Y. Xu. 2023. Ratio of histamine-producing/non-histamine-producing subgroups of *Tetragenococcus halophilus* determines the histamine accumulation during spontaneous fermentation of soy sauce. *Appl. Environ. Microbiol.* 89(3):e01884-22. <https://doi.org/10.1128/aem.01884-22>.
34. Molenaar, D., J. S. Bosscher, B. Ten Brink, A. J. Driessen, and W. N. Konings. 1993. Generation of a proton motive force by histidine decarboxylation and electrogenic histidine/histamine antiport in *Lactobacillus buchneri*. *J. Bacteriol.* 175:2864–2870. <https://doi.org/10.1128/jb.175.10.2864-2870.1993>.
35. Mou, Z., Y. Yang, A. B. Hall, and X. Jiang. 2021. The taxonomic distribution of histamine-secreting bacteria in the human gut microbiome. *BMC Genomics* 22:1–11. <https://doi.org/10.1186/s12864-021-08004-3>.
36. Naila, A., S. Flint, G. C. Fletcher, P. J. Bremer, and G. Meerdink. 2011. Biogenic amines and potential histamine-forming bacteria in rihaakuru (a cooked fish paste). *Food Chem.* 128(2):479–484. <https://doi.org/10.1016/j.foodchem.2011.03.057>.
37. Narkthewan, P., and W. Makkapan. 2020. Determination of histamine-forming bacteria, total coliform and *Escherichia coli* in fermented foods. *Geomate J.* 19(72):69–74. <https://doi.org/10.21660/2020.72.5695>.
38. Nevado, D. L., S. D. Santos, G. Bastian, J. Deyta, E. J. Managuelod, J. A. Fortaleza, and R. De Jesus. 2023. Detection, identification, and inactivation of histamine-forming bacteria in seafood: a mini-review. *J. Food Prot.* 100049. <https://doi.org/10.1016/j.jfp.2023.100049>.
39. Nor-Khaizura, M. A. R., H. Zaiton, B. Jamilah, R. A. G. Rusul, and M. R. Ismail-Fitry. 2009. Histamine and histamine-forming bacteria in keropok lekor (Malaysian fish sausage) during processing. *Food Sci. Tech. Res.* 15(4):395–402. <https://doi.org/10.3136/fstr.15.395>.
40. Oktariani, A. F., Y. Ramona, P. E. Sudaryatma, I. A. M. M. Dewi, and K. Shetty. 2022. Role of marine bacterial contaminants in histamine formation in seafood products: a review. *Microorganisms* 10:1197. <https://doi.org/10.3390/microorganisms10061197>.
41. Pasini, F., F. Soglia, M. Petracci, M. F. Caboni, S. Marziali, C. Montanari, F. Gardini, L. Grazia, and G. Tabanelli. 2018. Effect of fermentation with different lactic acid bacteria starter cultures on biogenic amine content and ripening patterns in dry fermented sausages. *Nutrients* 10(10):1497. <https://doi.org/10.3390/nu10101497>.
42. Podeur, G. 2014. Quantification des bactéries histaminogènes et maîtrise de la formation d’histamine dans les produits marins par biopréservation. Ph.D. thesis. Université de Nantes. Ecole doctorale N°495 Végétal, environnement, nutrition, agroalimentaire, mer. Available at: <https://archimer.ifremer.fr/doc/00241/35203/>. Accessed 22 May 2024.

43. Qasim, D. A. 2023. Isolation and identification of some types of histamine-producing bacteria from shrimp in local markets. *Bionatura* 1(8):1–10. <https://doi.org/10.21931/RB/CSS/S2023.08.01.47>.
44. Rachmawati, N., S. Powell, T. Ross, and M. Tamplin. 2023. Characterization of histamine-producing bacteria isolated during processing of Indonesian salted-boiled longtail (*Thunnus* spp.) and eastern little tuna (*Euthynnus* spp.). *J. Aquat. Food Prod. Tech.* 1–10. <https://doi.org/10.1080/10498850.2023.2219248>.
45. Raja, S., A. Alagiri, A. Ashok, J. Parthiban, and V. Meiyalagan. 2023. Isolation and identification of histamine-producing bacteria in frigate tuna *Auxis thazard*. *Kongunadu Res. J.* 10(2):15–20. <https://doi.org/10.26524/krij.2023.11>.
46. Rodriguez-Jerez, J. J., V. Giaccone, G. Colavita, and E. Parisi. 1994. *Bacillus macerans*—a new potent histamine producing micro-organism isolated from Italian cheese. *Food Microbiol.* 11(5):409–415. <https://doi.org/10.1006/fmic.1994.1046>.
47. Schloss, P. D., R. A. Girard, T. Martin, J. Edwards, and J. C. Thrash. 2016. Status of the archaeal and bacterial census: an update. *MBio* 7(3):10–1128. <https://doi.org/10.1128/mBio.00201-16>.
48. Simora, R. C., and E. M. Peralta. 2018. Occurrence of histamine and histamine-forming bacteria in Philippine traditional dried-salted fish products. *Asian Fish Sci.* 311:73–88. <https://doi.org/10.33997/j.afs.2018.31.2.001>.
49. Sitdhipol, J., S. Tanasupawat, P. Tepkasikul, P. Yukphan, A. Tosukhowong, T. Itoh, S. Benjakul, and W. Visessanguan. 2013. Identification and histamine formation of *Tetragenococcus* isolated from Thai fermented food products. *Ann. Microbiol.* 63:745–753. <https://doi.org/10.1007/s13213-012-0529-1>.
50. Stratton, J. E., R. W. Hutkins, and S. L. Taylor. 1991. Biogenic amines in cheese and other fermented foods: a review. *J. Food Prot.* 54(6):460–470. <https://doi.org/10.4315/0362-028X-54.6.460>.
51. Takai, J., H. Ohtsu, A. Sato, S. Uemura, T. Fujimura, M. Yamamoto, and T. Moriguchi. 2019. Lipopolysaccharide-induced expansion of histidine decarboxylase-expressing Ly6G+ myeloid cells identified by exploiting histidine decarboxylase BAC-GFP transgenic mice. *Sci. Rep.* 9(1):15603. <https://doi.org/10.1038/s41598-019-51716-6>.
52. Tembhurne, M., A. Ghag, H. Sanathkumar, and B. B. Nayak. 2013. Dominance of enterobacteria among histamine-producing bacteria isolated from Indian mackerel. *Adv. Microbiol.* 3(7):537–542. <http://dx.doi.org/10.4236/aim.2013.37072>.
53. Tham, W. 1988. Histamine formation by enterococci isolated from home-made goat cheeses. *Int. J. Food Microbiol.* 7(2):103–108. [https://doi.org/10.1016/0168-1605\(88\)90002-5](https://doi.org/10.1016/0168-1605(88)90002-5).
54. Tolmasky, M. E., L. A. Actis, and J. H. Crosa. 1995. A histidine decarboxylase gene encoded by the *Vibrio anguillarum* plasmid pJM1 is essential for virulence: histamine is a precursor in the biosynthesis of anguibactin. *Mol. Microbiol.* 15:87–95. <https://doi.org/10.1111/j.1365-2958.1995.tb02223.x>.
55. Torido, Y., H. Takahashi, T. Kuda, and B. Kimura. 2012. Analysis of the growth of histamine-producing bacteria and histamine accumulation in fish during storage at low temperatures. *Food Control* 26(1):174–177. <https://doi.org/10.1016/j.foodcont.2012.01.009>.
56. Tsai, Y. H., H. F. Kung, T. M. Lee, G. T. Lin, and D. F. Hwang. 2004. Histamine-related hygienic qualities and bacteria found in popular commercial scombroid fish fillets in Taiwan. *J. Food Prot.* 67(2):407–412. <https://doi.org/10.4315/0362-028X-67.2.407>.
57. Tsai, Y. H., H. F. Kung, Q. L. Lin, J. H. Hwang, S. H. Cheng, C. I. Wei, and D. F. Hwang. 2005. Occurrence of histamine and histamine-forming bacteria in kimchi products in Taiwan. *Food Chem.* 90(4):635–641. <https://doi.org/10.1016/j.foodchem.2004.04.024>.
58. Tsai, Y. H., C. Y. Lin, L. T. Chien, T. M. Lee, C. I. Wei, and D. F. Hwang. 2006. Histamine contents of fermented fish products in Taiwan and isolation of histamine-forming bacteria. *Food Chem.* 98(1):64–70. <https://doi.org/10.1016/j.foodchem.2005.04.036>.
59. Uehara, S., M. Kobayashi, K. Kimura, J. Suzuki, and K. Sadamasu. 2021. Rapid identification of histamine-producing bacteria isolated from fish using MALDI-TOF MS. *J. Food Sci. Tech.* 58:4055–4061. <https://doi.org/10.1007/s13197-021-05092-7>.
60. Visciano, P., M. Schirone, and A. Paparella. 2020. An overview of histamine and other biogenic amines in fish and fish products. *Foods* 9(12):1795. <https://doi.org/10.3390/foods9121795>.
61. Zhernov, Y. V., M. Y. Simanduyev, O. K. Zaostrovtsseva, E. E. Semeniako, K. I. Kolykhalova, I. A. Fadeeva, M. I. Kashutina, S. O. Vysochanskaya, E. V. Belova, D. V. Shcherbakov, V. A. Sukhov, E. A. Sidorova, and O. V. Mitrokhin. 2023. Molecular mechanisms of scombroid food poisoning. *Int. J. Mol. Sci.* 24(1):809. <https://doi.org/10.3390/ijms24010809>.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Acinetobacter baumannii</i>	Correct		Unique	Negative	Aerobic	No
<i>Acinetobacter hwoffii</i>	Correct		Unique	Negative	Aerobic	No
<i>Actinobacillus ureae</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Aeromonas hydrophila</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Aeromonas salmonicida</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Aeromonas sobria</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Aeromonas</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Alcaligenes faecalis</i>	Correct		Unique	Negative	Aerobic	No
<i>Alteromonas putrefaciens</i>	Synonym	<i>Shewanella putrefaciens</i>		Negative	Facultative anaerobic	No
<i>Bacillus amyloliquefaciens</i>	Correct		Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus cibi</i>	Synonym	<i>Metabacillus indicus</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus coagulans</i>	Synonym	<i>Heyndrickxia coagulans</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus firmus</i>	Synonym	<i>Cytobacillus firmus</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus flexus</i>	Synonym	<i>Priestia flexa</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus licheniformis</i>	Correct		Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus macerans</i>	Synonym	<i>Paenibacillus macerans</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus malacitensis</i>	Synonym	<i>Bacillus halotolerans</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus massiliensis</i>	Synonym	<i>Ureibacillus massiliensis</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus megaterium</i>	Synonym	<i>Priestia megaterium</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus oleronius</i>	Synonym	<i>Heyndrickxia oleronia</i>	Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus polyfermenticus</i>	Preferred name		Unique	Positive	Facultative anaerobic	Endospore
<i>Bacillus</i> spp.	Genus only			Positive	Facultative anaerobic	Endospore
<i>Bacillus subtilis</i>	Correct		Unique	Positive	Facultative anaerobic	Endospore

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Bacillus thuringiensis</i>	Correct		Unique	Positive	Facultative anaerobic	Endospore
<i>Brevibacillus brevis</i>	Correct		Unique	Positive	Aerobic	Endospore
<i>Brevibacterium casei</i>	Correct		Unique	Positive	Aerobic	No
<i>Citrobacter amalonaticus</i>	Correct		Unique	Negative	Aerobic	No
<i>Citrobacter braakii</i>	Correct		Unique	Negative	Aerobic	No
<i>Citrobacter farmeri</i>	Correct		Unique	Negative	Aerobic	No
<i>Citrobacter freundii</i>	Correct		Unique	Negative	Aerobic	No
<i>Citrobacter koseri</i>	Correct		Unique	Negative	Aerobic	No
<i>Citrobacter</i> spp.	Genus only			Negative	Aerobic	No
<i>Clostridium irregular</i>	Synonym	<i>Asaccharospora irregularis</i>	Unique	Positive	Anaerobic	Endospore
<i>Clostridium perfringens</i>	Correct		Unique	Positive	Anaerobic	Endospore
<i>Clostridium</i> spp.	Genus only			Positive	Anaerobic	Endospore
<i>Cronobacter sakazakii</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Edwardsiella hoshinae</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Enterobacter aerogenes</i>	Synonym	<i>Klebsiella aerogenes</i>		Negative	Facultative anaerobic	No
<i>Enterobacter amnigenus</i>	Synonym	<i>Lelliottia amnigena</i>	Unique	Negative	Facultative anaerobic	No
<i>Enterobacter cloacae</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Enterobacter gergovia</i>	Synonym	<i>Pluralibacter gergoviae</i>	Unique	Negative	Facultative anaerobic	No
<i>Enterobacter hormaechei</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Enterobacter</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Enterobacter taylorae</i>	Synonym	<i>Enterobacter cancerogenus</i>	Unique	Negative	Facultative anaerobic	No
<i>Enterobacter xianfengensis</i>	No BacDive		Unique	Negative	Facultative anaerobic	No
<i>Enterobacteriaceae</i>	Family			Negative	NA ^a	NA
<i>Enterococcus casseliflavus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Enterococcus faecalis</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Erwinia persicina</i>	Correct		Unique	Negative	Facultative anaerobic	No

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Escherichia coli</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Escherichia hermannii</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Hafnia alvei</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Hafnia paralvei</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Klebsiella aerogenes</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Klebsiella oxytoca</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Klebsiella pneumoniae</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Klebsiella pneumoniae</i> subsp. <i>pneumoniae</i>	Subsp.			Negative	Facultative anaerobic	No
<i>Klebsiella</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Klebsiella variicola</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Lactocaseibacillus casei</i>	Correct		Unique	Positive	Anaerobic	No
<i>Lactocaseibacillus paracasei</i>	Correct		Unique	Positive	Anaerobic	No
<i>Lactiplantibacillus plantarum</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Lactobacillus bavaricus</i>	Synonym	<i>Latilactobacillus sakei</i>		Positive	Aerotolerant	No
<i>Lactobacillus brevis</i>	Synonym	<i>Levilactobacillus brevis</i>		Positive	Aerotolerant	No
<i>Lactobacillus buchneri</i>	Synonym	<i>Lenticolactobacillus buchneri</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus curvatus</i>	Synonym	<i>Latilactobacillus curvatus</i>		Positive	Aerotolerant	No
<i>Lactobacillus delbrueckii</i>	Correct	<i>Lentilactobacillus hilgardii</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus delbrueckii</i> subsp. <i>lactis</i>	Subsp.			Positive	Aerotolerant	No
<i>Lactobacillus helveticus</i>	Correct		Unique	Positive	Aerotolerant	No
<i>Lactobacillus hilgardii</i>	Synonym	<i>Lentilactobacillus hilgardii</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus mali</i>	Synonym	<i>Liquorilactobacillus mali</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus para. paracasei</i>	Subsp.			Positive	Aerotolerant	No

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Lactobacillus parabuchneri</i>	Synonym	<i>Lentilactobacillus parabuchneri</i>	Positive	Aerotolerant	No	
<i>Lactobacillus paracasei</i>	Synonym	<i>Lactocaseibacillus paracasei</i>		Positive	Aerotolerant	No
<i>Lactobacillus paracollinoides</i>	Synonym	<i>Secundilactobacillus paracollinoides</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus pentosus</i>	Synonym	<i>Lactiplantibacillus pentosus</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus plantarum</i>	Synonym	<i>Lactiplantibacillus plantarum</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus rhamnosus</i>	Synonym	<i>Lactocaseibacillus rhamnosus</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus rossiae</i>	Synonym	<i>Furfurilactobacillus rossiae</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus saerimneri</i> 30a	Synonym	<i>Ligilactobacillus saerimneri</i>	Unique	Positive	Aerotolerant	No
<i>Lactobacillus</i> spp.	Genus only			Positive	Aerotolerant	No
<i>Lactococcus garvieae</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Lactococcus lactis</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Lactococcus lactis</i> subsp. <i>lactis</i>	Subsp.			Positive	Facultative anaerobic	No
<i>Latilactobacillus curvatus</i>	Correct		Unique	Positive	Microaerophile	No
<i>Latilactobacillus sakei</i>	Correct		Unique	Positive	Microaerophile	No
<i>Lenticolactobacillus buchneri</i>	No BacDive		Unique	Positive	Unknown	No
<i>Lenticolactobacillus parabuchneri</i>	No BacDive		Unique	Positive	Unknown	No
<i>Lentilactobacillus hilgardii</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Lentilactobacillus parabuchneri</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Leuconostoc mesenteroides</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Leuconostoc</i> spp.	Genus only			Positive	Facultative anaerobic	No
<i>Levilactobacillus brevis</i>	Correct		Unique	Positive	Anaerobic	No
<i>Limosilactobacillus reuteri</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Limosilactobacillus vaginalis</i>	Correct		Unique	Positive	Facultative anaerobic	No

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Listeria innocua</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Listonella anguillarum</i>	Correct		Unique	Negative	Aerobic	No
<i>Lysinibacillus boronitolerans</i>	Correct		Unique	Positive	Aerobic	Endospore
<i>Micrococcus</i> spp.	Genus only		Unique	Positive	Aerobic	No
<i>Morganella morganii</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Morganella psychrotolerans</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Morganella</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Neisseria zoodegmatis</i>	Correct		Unique	Negative	Aerobic	No
<i>Oenococcus oeni</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Pantoea agglomerans</i>	Correct		Unique	Negative	Aerobic	No
<i>Pantoea dispersa</i>	Correct		Unique	Negative	Aerobic	No
<i>Pantoea</i> spp.	Genus only			Negative	Aerobic	No
<i>Pediococcus damnosus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Pediococcus parvulus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Pediococcus pentosaceus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Photobacterium angustum</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium aquimaris</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium damsela</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium damsela</i> subsp. <i>damsela</i>	Subsp.			Negative	Facultative anaerobic	No
<i>Photobacterium iliopiscarium</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium kishitanii</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium leiognathi</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium phosphoreum</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Photobacterium psychrotolerans</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Plesiomonas shigelloides</i>	Correct		Unique	Negative	Facultative anaerobic	No

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Pluralibacter pyrinus</i>	Correct		Unique	Negative	Aerobic	No
<i>Proteus columbae</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Proteus marina</i>	No BacDive		Unique	Negative	Facultative anaerobic	No
<i>Proteus mirabilis</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Proteus penneri</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Proteus</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Proteus vulgaris</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Providencia rettgeri</i>	Correct		Unique	Negative	Facultative aerobic	No
<i>Providencia rustigianii</i>	Correct		Unique	Negative	Facultative aerobic	No
<i>Providencia stuartii</i>	Correct		Unique	Negative	Facultative aerobic	No
<i>Pseudomonas aeruginosa</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Pseudomonas fluorescens</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Pseudomonas putida</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Pseudomonas</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Pseudomonas syringae</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Psychrobacter pulmonis</i>	Correct		Unique	Negative	Aerobic	No
<i>Rahnella agnatilis</i>	No BacDive	<i>Rahnella aquatilis</i>				
Unique	Negative	<i>Facultative anaerobic</i>	No			
<i>Rahnella</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Raoultella ornithinolytica</i>	Synonym	<i>Klebsiella ornithinolytica</i>	Unique	Negative	Facultative anaerobic	No
<i>Raoultella planticola</i>	Synonym	<i>Klebsiella planticola</i>	Unique	Negative	Facultative anaerobic	No
<i>Raoultella</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Salmonella enterica</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Serratia fonticola</i>	Correct		Unique	Negative	Facultative anaerobic	No

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Serratia liquefaciens</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Serratia marcescens</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Serratia</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Shewanella baltica</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Shewanella haliotis</i>	Synonym	<i>Shewanella algae</i>	Unique	Negative	Facultative anaerobic	No
<i>Shewanella putrefaciens</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Shewanella</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Shigella flexneri</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Sphingomonas paucimobilis</i>	Correct		Unique	Negative	Aerobic	No
<i>Staphylococcus aureus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus capitis</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus epidermidis</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus hominis</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus intermedius</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus kloosii</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus pasteurii</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus saprophyticus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus sciuri</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Staphylococcus</i> spp.	Genus only			Positive	Facultative anaerobic	No
<i>Staphylococcus xylosus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Streptococcus faecalis</i>	Synonym	<i>Enterococcus faecalis</i>		Positive	Facultative anaerobic	No
<i>Streptococcus faecium</i>	Synonym	<i>Enterococcus faecium</i>	Unique	Positive	Facultative anaerobic	No

Table continued on the next page.

APPENDIX A. Histamine-forming bacteria found in the literature that are Gram-positive or Gram-negative, their oxygen requirements, and their spore-forming ability (cont.)

Bacterial species named in text	Taxonomic status	Correct or alternate name	Unique	Gram-positive or Gram-negative	Environment	Spores
<i>Streptococcus thermophilus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Tetragenococcus halophilus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Tetragenococcus muriaticus</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Tetragenococcus</i> spp.	Genus only			Positive	Facultative anaerobic	No
<i>Vibrio alginolyticus</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Vibrio anguillarum</i>	Synonym	<i>Listonella anguillarum</i>		Negative	Facultative anaerobic	No
<i>Vibrio fluvialis</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Vibrio halodenitrificans</i>	No BacDive		Unique	Negative	Facultative anaerobic	No
<i>Vibrio harveyi</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Vibrio mimicus</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Vibrio olivaceus</i>	No BacDive		Unique	Negative	Facultative anaerobic	No
<i>Vibrio parahaemolyticus</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Vibrio</i> spp.	Genus only			Negative	Facultative anaerobic	No
<i>Vibrio vulnificus</i>	Correct		Unique	Negative	Facultative anaerobic	No
<i>Weissella cibaria</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Weissella confusa</i>	Correct		Unique	Positive	Facultative anaerobic	No
<i>Weissella paramesenteroides</i>	Correct		Unique	Positive	Facultative anaerobic	No

^aNA, not applicable.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number

Bacterial species	Author, y	Ref no.	DOI no.
<i>Acinetobacter baumannii</i>	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
<i>Acinetobacter lwoffii</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Actinobacillus ureae</i>	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
<i>Aeromonas hydrophila</i>	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Lim, 2016	(32)	https://doi.org/10.1186/s41240-016-0040-x
<i>Aeromonas salmonicida</i>	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
<i>Aeromonas sobria</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Aeromonas</i> spp.	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Alcaligenes faecalis</i>	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
<i>Alteromonas putrefaciens</i>	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
<i>Bacillus amyloliquefaciens</i>	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
<i>Bacillus cibi</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus coagulans</i>	Tsai et al., 2006	(58)	https://doi.org/10.1016/j.foodchem.2005.04.036
<i>Bacillus firmus</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus flexus</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus licheniformis</i>	Lim, 2016	(32)	https://doi.org/10.1186/s41240-016-0040-x
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Bacillus macerans</i>	Rodriguez-Jerez et al., 1994	(46)	https://doi.org/10.1006/fmic.1994.1046
<i>Bacillus malacitensis</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus massiliensis</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus megaterium</i>	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
	Tsai et al., 2006	(58)	https://doi.org/10.1016/j.foodchem.2005.04.036
<i>Bacillus oleronius</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus polyfermenticus</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Bacillus</i> spp.	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001
<i>Bacillus subtilis</i>	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Bacillus thuringiensis</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Brevibacillus brevis</i>	Tsai et al., 2005	(57)	https://doi.org/10.1016/j.foodchem.2004.04.024
<i>Brevibacterium casei</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Citrobacter amalonaticus</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Citrobacter braakii</i>	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Citrobacter farmeri</i>	Narkthewan & Makkapan, 2020	(37)	https://doi.org/10.21660/2020.72.5695
<i>Citrobacter freundii</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsv51i2.10819
	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Citrobacter koseri</i>	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
<i>Citrobacter</i> spp.	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
<i>Clostridium irregular</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Clostridium perfringens</i>	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Clostridium</i> spp.	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Cronobacter sakazakii</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Edwardsiella hoshinae</i>	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
<i>Enterobacter aerogenes</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Bjornsdottir-Butler et al., 2015	(6)	https://doi.org/10.4315/0362-028X.JFP-15-012
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
	Tsai et al., 2005	(57)	https://doi.org/10.4315/0362-028X-68.8.1690
<i>Enterobacter amnigenus</i>	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Enterobacter cloacae</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Enterobacter gergovia</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
<i>Enterobacter hormaechei</i>	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
<i>Enterobacter</i> spp.	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
<i>Enterobacter taylorae</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Enterobacter xianfengensis</i>	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
<i>Enterobacteriaceae</i>	Lavizzari et al., 2010	(31)	https://doi.org/10.4315/0362-028X-73.2.385
<i>Enterococcus casseliflavus</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Enterococcus faecalis</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Erwinia persicina</i>	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
<i>Escherichia coli</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Escherichia hermannii</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
<i>Hafnia alvei</i>	Bang et al., 2009	(3)	https://doi.org/10.5352/JLS.2009.19.2.277
	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Lavizzari et al., 2010	(31)	https://doi.org/10.4315/0362-028X-73.2.385
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Hafnia paralvei</i>	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
<i>Klebsiella aerogenes</i>	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Klebsiella oxytoca</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
<i>Klebsiella pneumoniae</i>	Bang et al., 2009	(3)	https://doi.org/10.5352/JLS.2009.19.2.277
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Klebsiella pneumoniae</i> subsp. <i>pneumoniae</i>	Lavizzari et al., 2010	(31)	https://doi.org/10.4315/0362-028X-73.2.385
<i>Klebsiella</i> spp.	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
<i>Klebsiella variicola</i>	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
<i>Lactocaseibacillus casei</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lactocaseibacillus paracasei</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lactiplantibacillus plantarum</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lactobacillus bavaricus</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Lactobacillus brevis</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Kung et al., 2005	(29)	https://doi.org/10.38212/2224-6614.2557
<i>Lactobacillus buchneri</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Fröhlich-Wyder et al., 2013	(21)	https://doi.org/10.1016/j.idairyj.2013.03.004
	Stratton et al., 1991	(50)	https://doi.org/10.4315/0362-028X-54.6.460
<i>Lactobacillus curvatus</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Kung et al., 2005	(29)	https://doi.org/10.38212/2224-6614.2557
<i>Lactobacillus delbrueckii</i> subsp. <i>lactis</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lactobacillus helveticus</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Lactobacillus hilgardii</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Lactobacillus mali</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Lactobacillus para. paracasei</i>	Kung et al., 2005	(29)	https://doi.org/10.38212/2224-6614.2557
<i>Lactobacillus parabuchneri</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Fröhlich-Wyder et al., 2013	(21)	https://doi.org/10.1016/j.idairyj.2013.03.004
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Lactobacillus paracasei</i>	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Lactobacillus paracollinoides</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Lactobacillus pentosus</i>	Kung et al., 2005	(29)	https://doi.org/10.38212/2224-6614.2557
<i>Lactobacillus plantarum</i>	Kung et al., 2005	(29)	https://doi.org/10.38212/2224-6614.2557
<i>Lactobacillus rhamnosus</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Lactobacillus rossiae</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Lactobacillus saerimneri</i> 30a	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Lactobacillus</i> spp.	Kung et al., 2005	(29)	https://doi.org/10.38212/2224-6614.2557
<i>Lactococcus garvieae</i>	Bang et al., 2009	(3)	https://doi.org/10.5352/JLS.2009.19.2.277
<i>Lactococcus lactis</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lactococcus lactis</i> subsp. <i>lactis</i>	Bang et al., 2009	(3)	https://doi.org/10.5352/JLS.2009.19.2.277
<i>Latilactobacillus curvatus</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Latilactobacillus sakei</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lentilactobacillus buchneri</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lentilactobacillus parabuchneri</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lentilactobacillus hilgardii</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Lentilactobacillus parabuchneri</i>	Botello-Morte et al., 2022	(7)	https://doi.org/10.1016/j.foodcont.2021.108595
<i>Leuconostoc mesenteroides</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Leuconostoc</i> spp.	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Levilactobacillus brevis</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Limosilactobacillus reuteri</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Limosilactobacillus vaginalis</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Listeria innocua</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
<i>Listonella anguillarum</i>	Podeur, 2014	(42)	https://archimer.ifremer.fr/doc/00241/35203/
<i>Lysinibacillus boronitolerans</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Micrococcus</i> spp.	Nor-Khaizura et al., 2009	(39)	https://doi.org/10.3136/fstr.15.395
	Bang et al., 2009	(3)	https://doi.org/10.5352/JLS.2009.19.2.277
	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Bjornsdottir-Butler et al., 2015	(6)	https://doi.org/10.4315/0362-028X.JFP-15-012
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsr.v51i2.10819
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Lavizzari et al., 2010	(31)	https://doi.org/10.4315/0362-028X-73.2.385

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Morganella morganii</i>	Lim, 2016	(32)	https://doi.org/10.1186/s41240-016-0040-x
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Uehara et al., 2021	(59)	https://doi.org/10.1007/s13197-021-05092-7
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Morganella psychrotolerans</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
<i>Morganella</i> spp.	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
<i>Neisseria zoodegmatidis</i>	Raja et al., 2023	(45)	https://doi.org/10.26524/krij.2023.11
<i>Oenococcus oeni</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Pantoea agglomerans</i>	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
<i>Pantoea dispersa</i>	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
<i>Pantoea</i> spp.	Lavizzari et al., 2010	(31)	https://doi.org/10.4315/0362-028X-73.2.385
<i>Pediococcus damnosus</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Pediococcus parvulus</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Pediococcus pentosaceus</i>	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Photobacterium angustum</i>	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
<i>Photobacterium aquimaris</i>	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Photobacterium damsela</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Uehara et al., 2021	(59)	https://doi.org/10.1007/s13197-021-05092-7
<i>Photobacterium damsela</i> subsp. <i>damsela</i>	Bjornsdottir-Butler et al., 2015	(6)	https://doi.org/10.4315/0362-028X.JFP-15-012
<i>Photobacterium iliopiscarium</i>	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Torido et al., 2012	(55)	https://doi.org/10.1016/j.foodcont.2012.01.009
<i>Photobacterium kishitanii</i>	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Photobacterium leiognathi</i>	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Photobacterium phosphoreum</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Torido et al., 2012	(55)	https://doi.org/10.1016/j.foodcont.2012.01.009
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Photobacterium psychrotolerans</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
<i>Plesiomonas shigelloides</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Pluralibacter pyrinus</i>	Uehara et al., 2021	(59)	https://doi.org/10.1007/s13197-021-05092-7
<i>Proteus columbae</i>	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
<i>Proteus marina</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Proteus mirabilis</i>	Ababouch et al., 1991	(1)	https://doi.org/10.1016/0740-0020(91)90005-M
	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
<i>Proteus penneri</i>	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
<i>Proteus spp.</i>	Ababouch et al., 1991	(1)	https://doi.org/10.1016/0740-0020(91)90005-M
<i>Proteus vulgaris</i>	Ababouch et al., 1991	(1)	https://doi.org/10.1016/0740-0020(91)90005-M
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	DeBeer et al., 2021	(13)	https://doi.org/10.4315/1541-9576-41.1.36
	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
	Raja et al., 2023	(45)	https://doi.org/10.26524/krj.2023.11
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
<i>Providencia rettgeri</i>	Ababouch et al., 1991	(1)	https://doi.org/10.1016/0740-0020(91)90005-M

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Providencia rustigianii</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Raja et al., 2023	(45)	https://doi.org/10.26524/krj.2023.11
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
<i>Providencia stuartii</i>	Ababouch et al., 1991	(1)	https://doi.org/10.1016/0740-0020(91)90005-M
<i>Pseudomonas aeruginosa</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Pseudomonas fluorescens</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Pseudomonas putida</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001
<i>Pseudomonas</i> spp.	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
	Nor-Khaizura et al., 2009	(39)	https://doi.org/10.3136/fstr.15.395
<i>Pseudomonas syringae</i>	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001
<i>Psychrobacter pulmonis</i>	Devivilla et al., 2019	(14)	https://doi.org/10.4315/0362-028X.JFP-19-101
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
<i>Rahnella agnatilis</i>	Tsai et al., 2004	(56)	https://doi.org/10.4315/0362-028X-67.2.407
<i>Rahnella</i> spp.	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
<i>Raoultella ornithinolytica</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Uehara et al., 2021	(59)	https://doi.org/10.1007/s13197-021-05092-7
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Raoultella planticola</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Bjornsdottir-Butler et al., 2015	(6)	https://doi.org/10.4315/0362-028X.JFP-15-012
	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Zhernov et al., 2023	(61)	https://doi.org/10.3390/ijms24010809
<i>Raoultella</i> spp.	Firth et al., 2023	(20)	https://doi.org/10.1128/spectrum.04720-22
<i>Salmonella enterica</i>	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Serratia fonticola</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Serratia liquefaciens</i>	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
<i>Serratia marcescens</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Lim, 2016	(32)	https://doi.org/10.1186/s41240-016-0040-x
	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Serratia</i> spp.	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
<i>Shewanella baltica</i>	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
<i>Shewanella haliotis</i>	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
<i>Shewanella putrefaciens</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
<i>Shewanella</i> spp.	Ginigaddarage et al., 2023	(22)	http://dx.doi.org/10.4038/jnsfsrv51i2.10819
<i>Shigella flexneri</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Rachmawati et al., 2023	(44)	https://doi.org/10.1080/10498850.2023.2219248
<i>Sphingomonas paucimobilis</i>	Raja et al., 2023	(45)	https://doi.org/10.26524/krj.2023.11
<i>Staphylococcus aureus</i>	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001
<i>Staphylococcus capitis</i>	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
	Tembhurne et al., 2013	(52)	http://dx.doi.org/10.4236/aim.2013.37072
<i>Staphylococcus epidermidis</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Staphylococcus hominis</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Staphylococcus intermedius</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Staphylococcus kloosii</i>	Narkthewan & Makkapan, 2020	(37)	https://doi.org/10.21660/2020.72.5695
<i>Staphylococcus pasteurii</i>	Kung et al., 2006	(27)	https://doi.org/10.1016/j.foodchem.2005.08.025
	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
<i>Staphylococcus saprophyticus</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Staphylococcus sciuri</i>	Kung et al., 2012	(28)	https://doi.org/10.1016/j.jfda.2014.10.009
<i>Staphylococcus</i> spp.	Nor-Khaizura et al., 2009	(39)	https://doi.org/10.3136/fstr.15.395
<i>Staphylococcus xylosum</i>	Kung et al., 2015	(26)	https://doi.org/10.1016/j.jfda.2014.10.009
	Lim, 2016	(32)	https://doi.org/10.1186/s41240-016-0040-x
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
<i>Streptococcus faecalis</i>	Tham, 1988	(53)	https://doi.org/10.1016/0168-1605(88)90002-5
<i>Streptococcus faecium</i>	Tham, 1988	(53)	https://doi.org/10.1016/0168-1605(88)90002-5
<i>Streptococcus thermophilus</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Calles-Enríquez et al., 2010	(8)	https://doi.org/10.1128/AEM.00827-10
	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Tetragenococcus halophilus</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Botello-Morte et al., 2022	(7)	https://doi.org/10.1016/j.foodcont.2021.108595

Table continued on the next page.

APPENDIX B. Histamine-forming bacteria, author and year, reference number, and DOI number (cont.)

Bacterial species	Author, y	Ref no.	DOI no.
<i>Tetragenococcus halophilus</i>	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
	Ma et al., 2023	(33)	https://doi.org/10.1128/aem.01884-22
	Sitdhipol et al., 2013	(49)	https://doi.org/10.1007/s13213-012-0529-1
<i>Tetragenococcus muriaticus</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
	Sitdhipol et al., 2013	(49)	https://doi.org/10.1007/s13213-012-0529-1
<i>Tetragenococcus</i> spp.	Comas-Basté et al., 2019	(10)	https://doi.org/10.5772/intechopen.84333
	Ferrante & Mercogliano, 2023	(18)	https://doi.org/10.1016/j.foodchem.2023.136046
<i>Vibrio alginolyticus</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Nevado et al., 2023	(38)	https://doi.org/10.1016/j.jfp.2023.100049
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001
<i>Vibrio anguillarum</i>	Tolmasky et al., 1995	(54)	https://doi.org/10.1111/j.1365-2958.1995.tb02223.x
<i>Vibrio fluvialis</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
<i>Vibrio halodenitrificans</i>	Naila et al., 2011	(36)	https://doi.org/10.1016/j.foodchem.2011.03.057
<i>Vibrio harveyi</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Vibrio mimicus</i>	Bjornsdottir et al., 2009	(5)	https://doi.org/10.4315/0362-028X-72.9.1987
<i>Vibrio olivaceus</i>	Bang et al., 2009	(3)	https://doi.org/10.5352/JLS.2009.19.2.277
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
<i>Vibrio parahaemolyticus</i>	Kim et al., 2001	(24)	https://doi.org/10.4315/0362-028X-64.10.1556
	Oktariani et al., 2022	(40)	https://doi.org/10.3390/microorganisms10061197
	Simora & Peralta, 2018	(48)	https://doi.org/10.33997/j.afs.2018.31.2.001
<i>Vibrio</i> spp.	Ababouch et al., 1991	(1)	https://doi.org/10.1016/0740-0020(91)90005-M
<i>Vibrio vulnificus</i>	Qasim, 2023	(43)	https://doi.org/10.21931/RB/CSS/S2023.08.01.47
<i>Weissella cibaria</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Weissella confusa</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017
<i>Weissella paramesenteroides</i>	Barbieri et al., 2019	(4)	https://doi.org/10.3390/foods8010017