

Molecular-based Surveillance in Food Manufacturing Facilities Using Next Generation Sequencing Techniques and Software

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BACKGROUND

Background



Phylogenetic Profiles of In-House Microflora in Drains at a Food Production Facility: Comparison and Biocontrol Implications of *Listeria*-Positive and -Negative Bacterial Populations

Edward Fox, Katie Solomon, John Moore, Patrick Wall and **Séamus Fanning**

Applied and Environmental Microbiology, **2014**, **80(11):3369**

Fox et al.

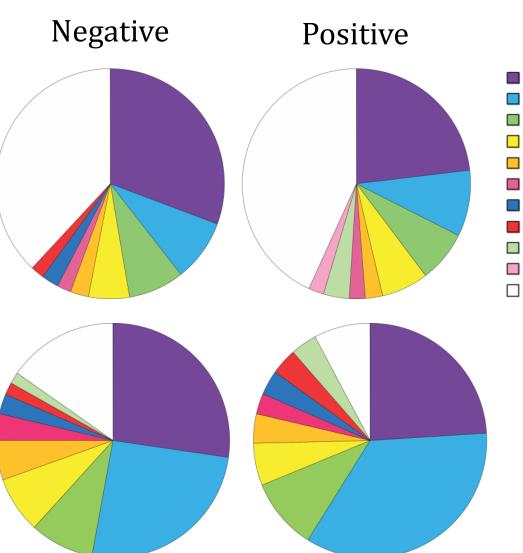


Study

- Examined interaction between Listeria species and other microorganisms
- Data gathered from drains in a meat production facility
- Four drains examined two Listeria positive, two Listeria negative
- Microbiome of drains examined, and overall diversity analysed



Microbiome Analysis



Lachnospiraceae Pseudomonadaceae Rikenellaceae Enterobacteriaceae Halomonadaceae ■ Bacillaceae Ruminococcaceae Prevotellaceae □ Flavobacteriaceae Comamonadaceae □ Unclassified and Others

Gammaproteobacteria

□ Unclassified and others

Clostridia Bacteroidia

Actinobacteria ■ Fusobacteria

Bacilli

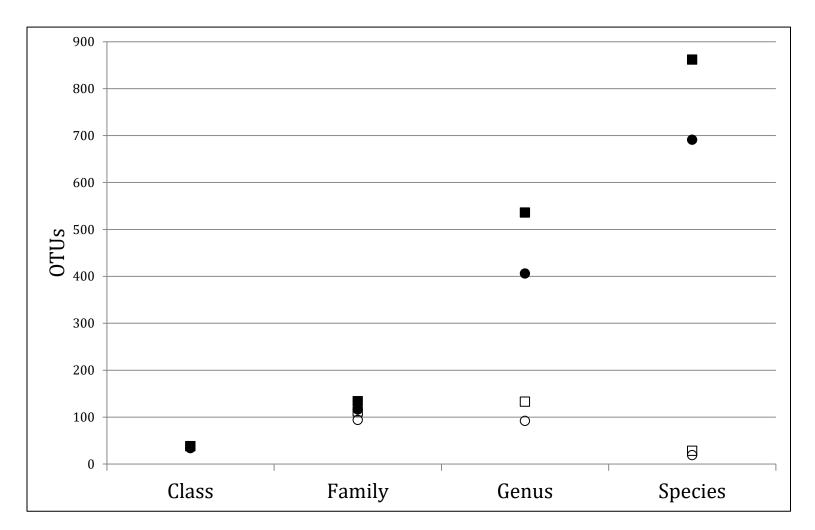
Classes

Alphaproteobacteria **Families** Deltaproteobacteria Betaproteobacteria



Fox et al.





•: *Listeria*-negative drains, overall diversity; ○: *Listeria*-negative drains, classified OTUs only; ■: *Listeria*-positive drains, overall diversity; □: *Listeria*-positive drains, classified OTUs only.

Results



Microbiome

- 21 families found *only* in Listeria positive drains
- Janthinobacterium, Prevotella, and Pseudomonas all associated with higher abundance in Listerianegative drains

Co-Culturing

- Presence of J. *Lividum* reduced the attachment and biofilm growth of L. *monocytogenes*
- E. *Gallinarum* caused significant increase in L. *monocytogenes* attachment and biofilm formation

Suggested Control Strategy



- Identify drains colonised by *Listeria*, and drains free of *Listeria*
- Determine microbiome and profile the local microbial communities
- Determine potential protagonists and antagonists

2

3

4

5

6

- Isolate key in-house protagonists and antagonists
- Examine the effect of these strains on *Listeria* attachment and biofilm formation
- Seed drains with non-pathogenic 'in-house' bacterial species that show inhibition of *Listeria* colonisation

Questions



- Can we quantify the relationship between the microbiome of a manufacturing facility and specific bacteria of interest?
- Can we quantify/predict what would be an effective biocontrol strategy?
- Can we examine specific bacteria using WGS in parallel with the microbiome?
- What traits can be identified using WGS and monitored over time and space?



THE S.A.F.E. PROJECT

The SAFE Project



- <u>Sequencing Alliance for Food</u>
 <u>Environments</u>
- Three year project in collaboration with:
 - University College Dublin Centre for Food Safety (UCD-CFS)
 - Five postdoctoral researchers
 - Creme Global
 - Six industry partners

Project Goals



Develop a pipeline of data using genomebased technology (WGS & WMS)

Develop mathematical models of safety and quality

Link gene-based information to phenotype for tracking/tracing of bacteria of interest

Use phenotypic data to highlight potential solution strategies to food safety and quality

Provide software tools to improve food safety & quality decision making steps

Tasks



Task 1

• Sampling plan design

Task 2

- Whole Genome Sequencing
- Microbiome Characterisation

Task 3

• Bioinformatic analysis

Task 4

• Production solution scenarios

Task 5

• Modelling and Software

Industry Partners











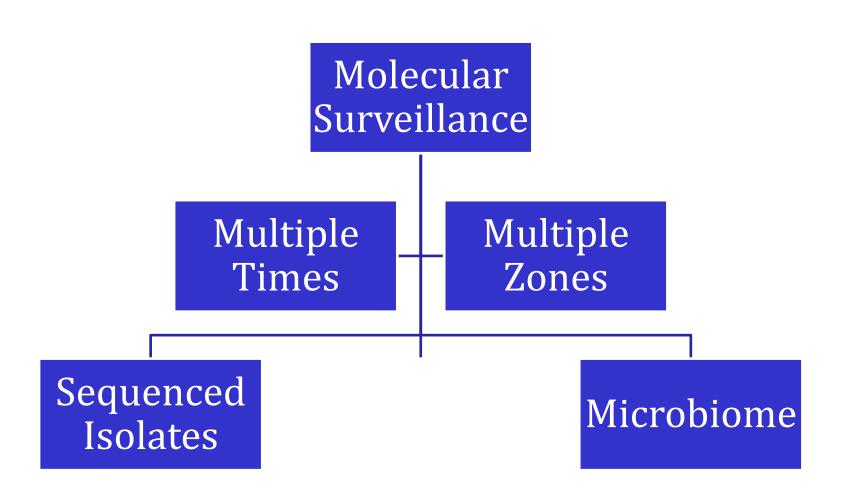




GREAT INGREDIENTS GREAT FOOD.

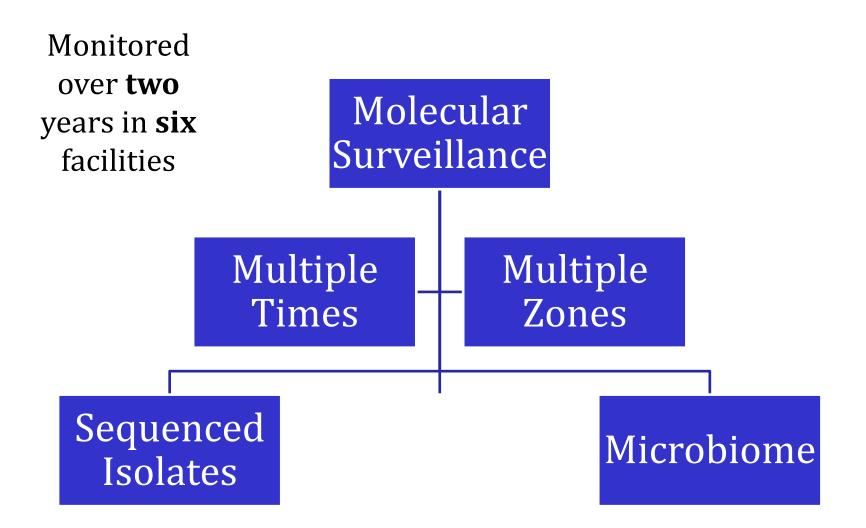
Sampling Plan





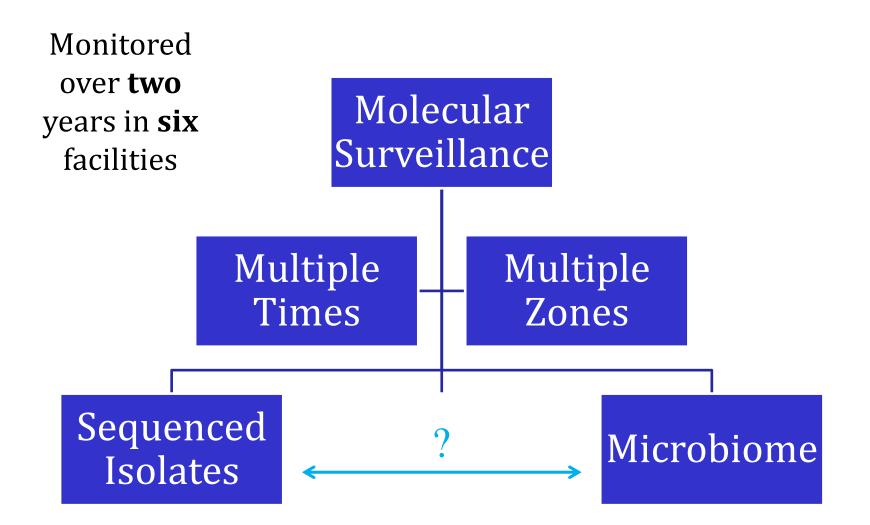
Sampling Plan





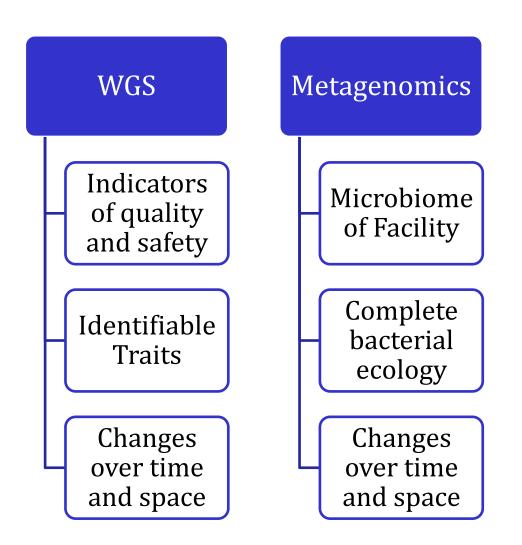
Sampling Plan





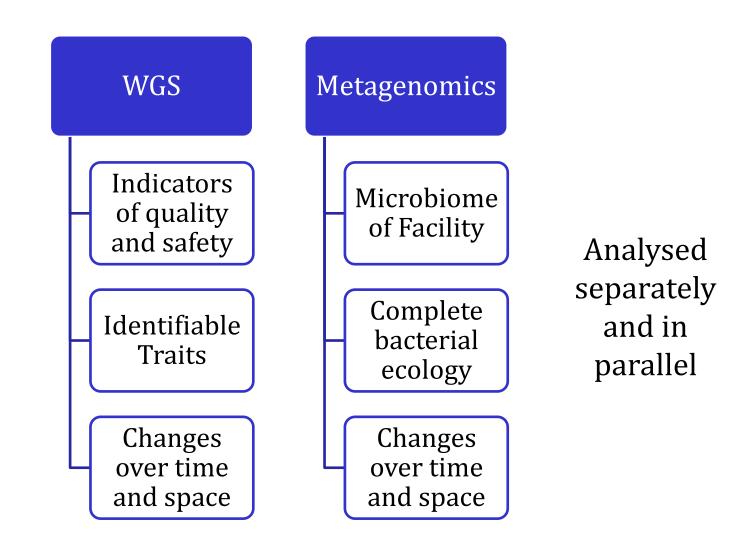
Sampling Data



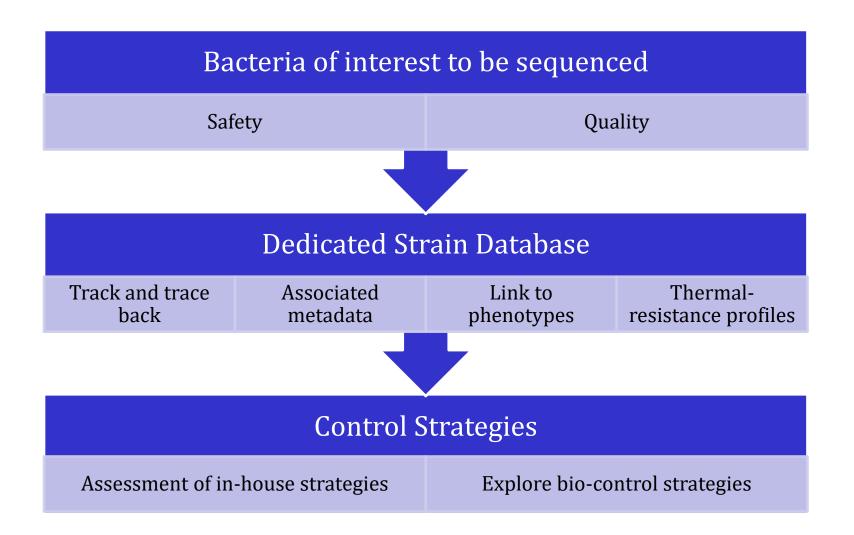


Sampling Data



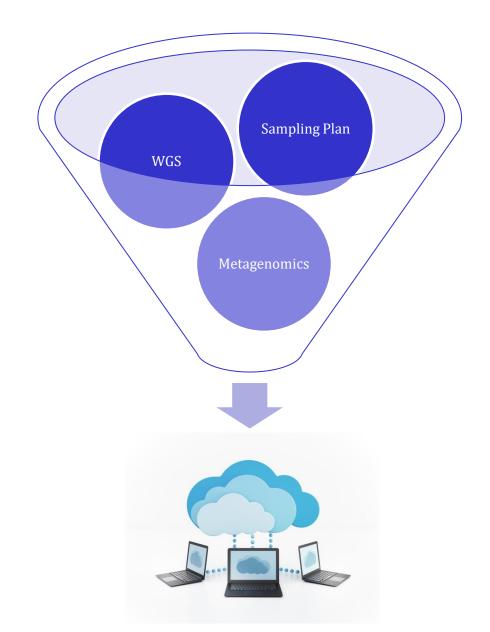






Final Output







SOFTWARE PLATFORM

SAFE

EMAIL

PASSWORD

LOGIN

Data > 2017 > Samples

Samples

Year	Month	Area	Sample Number
2017	Feb	Dry room	12345
2017	Feb	Wet room	22345
2017	Mar	Wet room	32345

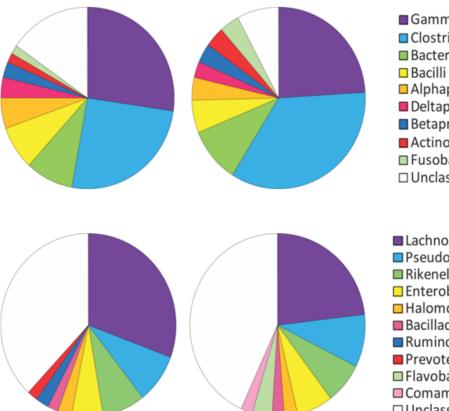
Data > 2017 > Samples

WGS Analysis

Isolate ID	Date	Location
10-00001	Feb-17	Dry Zone, High Risk
10-00002	Feb-17	Wet Zone, Low Risk
10-00008	Feb-17	Dry Zone, High Risk
10-00009	Feb-17	Wet Zone, Low Risk
10-00017	Feb-17	Dry Zone, High Risk
11-00005	Jun-17	Wet Zone, Low Risk
11-00537	Jun-17	Wet Zone, Low Risk
11-00538	Jun-17	Wet Zone, Low Risk
11-01558	Jun-17	Wet Zone, Low Risk
11-01559	Jun-17	Wet Zone, Low Risk

AM Resistance	Virulence	Thermal Resistance	Biofilm Formation	Biocide

Sample 123



Gammaproteobacteria
 Clostridia
 Bacteroidia
 Bacilli
 Alphaproteobacteria
 Deltaproteobacteria
 Betaproteobacteria
 Actinobacteria
 Fusobacteria
 Unclassified and others

Lachnospiraceae
 Pseudomonadaceae
 Rikenellaceae
 Enterobacteriaceae
 Halomonadaceae
 Bacillaceae
 Ruminococcaceae
 Prevotellaceae
 Flavobacteriaceae
 Comamonadaceae
 Unclassified and Others



Thank You

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